P. Sindhumole, P.Manju and Vijayaragahvakumar

(L.) Moench). Ph.D (Hort.) thesis, University of Agricultural Sciences, Bangalore, p. 147.

- Sheela, M.N. (1994). Induction of genetic recombination in interspecific crosses of Abelmoschus. Ph.D thesis, Kerala Agricultural University, Thrissur, p. 182.
- Singh, R.K. and Choudhary, B.D. (1985). Biometrical Methods in Quantitative Genetic Analysis. Kalyani Publishers, New Delhi, p.304.
- Singh, K., Malik, Y., Kalloo, S. and Mehrotra, N. (1974). Genetic variability and correlation studies in bhindi (Abelmoschus esculentus (L.) Moench). Veg. Sci. 1: 47-54.
- Thaker, D.N., Tikka, S.B.S., Patel, K.K. andUkani, S.J. (1981). Analysis of parameters of variability in okra. (Abelmoschus esculentus (L.) Moench). Indian J. Hort. 38: 232-235.
- Yassin, G.M. and Anbu, S. (1997). Variability studies in bhendi. South Indian Hort. 45: 13-15.
- Yadav, D.S. (1986). Variability and interrelations between yield and its components in okra (Abelmoschus esculentus (L.) Moench). Indian J. Hort. 43: 274-277.

Madras Agric. J., 93 (7-12) : 266-268 July-December 2006

https://doi.org/10.29321/MAJ.10.100764

Research Notes

High frequency of callusing from root explants of Assam Rice Collection

S. CHITRA AND C.R. ANANDA KUMAR

Dept. of Plant breeding and Genetics, Agricultural College and Res. Instt, Killikulam - 628 252, Tamil Nadu, India.

The regeneration of plants from cell and tissue culture is an important and essential component of biotechnology that is required for the genetic manipulation and improvement of plants. Rice is the world's single most important food crop and a primary food source for more than one third of world's population. Callusing and subsequent regeneration of rice from different explants were reported by Rueb *et al.* (1994), Seraj *et al.* (1997), Wenjing *et al.* (1997) and Mukopadhyay *et al.* (1997). However there are only a few reports on the induction of callus from root explants in rice. Mature seeds of Assam rice collection were obtained from Central Rice Research

Institute Cuttack, along with *japonica* check Taipei 309, and were used for this study. For obtaining root explants, mature seeds were dehusked and sterilized under aseptic condition. MS medium was supplemented with 2 mgL-1 1AA or NAA (Indole acetic acid or Naphthalene acetic acid) and used for obtaining adventitious roots from rice seedlings. Eighteen to twenty days old roots from rice seedlings were selected as explants (Anju John and Prathapasenan, 1999). Effects of different carbon sources (sucrose/maltose) were also tested. Approximately 1.5 cm length root explants were transferred to callus induction medium (MS medium containing 2 mg 2, 4-D L⁻¹).

266



268

Callus formation occurred after 10 days when cultures were incubated in dark at 25+2°C for 3 days and then kept in the light room under 16 hr light and 8 hr dark photoperiod. Callus was developed from cut ends of root explants and adventitious root tips only (Anju John and Prathapasenan, 1999). Even within the indica group, there were significant variations in the in vitro culture response among different genotypes (Hoque and Mansfield, 2004). Among the ARC lines, ARC 15759 expressed high frequency of callus induction (96.6%) than check Taipei 309 (86.7%). The lines ARC 18023 and ARC 18214 were at par with check (Fig. 1). High frequency callus induction was obtained when using maltose as carbon source instead of using sucrose (Fig. 2).

References

Anju,J. and Prathapasenan, G. (1999). High frequency of plantlet regeneration from root explants of rice (Oryza sativa L.) var CSR -10. Phytomorphology, 49: 165-169.

- Hoque, M.E.and Mansfield, J.W. (2004). Effect of genotype and explant age on callus induction and subsequent plant regeneration from root derived callus of indica rice genotypes. *Pl. Cell. Tiss. Org. Cult.* **78(3):** 217-223.
- Mukhopadhyay, A., Minhas, D. and Grover, A. (1997). Callusing from root explants. Adventitious root formation proceeds callus initiation response. *Curr, Sci.*, **13:** 465-469.
- Rueb, S., Leneman, M., Schilperoot, R.A. and Hensgens, A.M. (1994). Efficient plant regeneration through somatic embryogenesis from callus induced on mature rice embryos (*Oryza sativa L.*) *Pl. Cell Tiss. Org Cult.*, **36:** 361-365.
- Seraj, Z.I., Islam, Z., Faruque, M.O., Devi, T. and Ahmed, S. (1997). Identification of regeneration potential of embryos derived calluses from various *indica* rice varieties. *Pl. Cell. Tiss. Org. Cult*, **48**: 9-13.
- Wenjing, T., Bao, L. and Miao, X. (1997). Establishing *japonica* suspensions retaining a high regeneration potential after 14 month of culture: *Pl. Cell. Tiss. Org. Cult.*, 47: 213-216.

Madras Agric. J., 93 (7-12) : 268-271 July-December 2006

Research Notes

Organic sources of nutrients on groundnut seed production

V. PARAMASIVAM, P. BALAMURUGAN AND S. MOHANDAS

Krishi Vigyan Kendra, Tindivanam, Tamil Nadu Agricultural University, Coimbatore - 641 003

India ranks first in groundnut acreage (6.4 million ha), which accounts for 23.87 per cent of the world total groundnut area. The average productivity of groundnut in India is 1125 kg ha⁻¹, which is far below the world's

average of 1449 kg ha⁻¹ (www.agrico.op.nic.in). In the wake of serious pollution problems and bio-magnification of toxic chemicals in the various biological systems, 'organic farming' is the right approach in the present day