Effect of foliar spray of nutrients and plant growth regulators (PGRs) for yield maximization in blackgram

N.SRITHARAN, ANITHA ARAVAZHI AND MALLIKA VANANGAMUDI

Department of Crop Physiology Tamil Nadu Agricultural University, Coimbatore - 641 003.

Abstract: An experiment was conducted during 2002 to study the morphological physiological and biochemical effects of foliar spray of nutrients and plant growth regulators on yield and productivity of blackgram. The treatments include foliar spray of 2% DAP, 0.2% Boric acid, 0.5% FeSO₄, 0.5% ZnSO₄, 0.5% sodium molybdate, 2% urea, 0.1% humic acid, 1% KC1, Salicylic acid 100 ppm, Brassinolide 0.1 ppm and humic acid 20 kg basal application. Among these treatments, foliar spray of 2% urea recorded the highest yield of 955.2 kg/ha followed by foliar spray of KC1 1% along with soil application of humic acid @ 20 kg/ha (926.2 kg/ha). The yield enhancement may be due to improved morphological, physiological and biochemical parameters. The tested chemicals particularly 2% urea were found effective towards yield maximization in black gram.

Introduction

Hormones play an important role on the vegetative and reproductive growth of any plant. At present, several synthetic hormones are employed by the farmers to boost the growth in the vegetative phase and subsequently to increase the yield. Several efforts have been made to increase the yield potential through breeding for high yielding varieties incorporating efficient irrigating methods and application of fertilizers, biocides and hormones. Nutrients such as DAP, N as urea, FeSO4, ZnSO4 are known to significantly influence the yield of pulses (Sujatha, 2001). Salicylic acid, BR and humic acid proved their efficiency in increasing germination (Senthilkumar, 2001) and seed yield in groundnut (Amal, 2001). In grain legumes including blackgram, the application of PGRs and nutrients decreases the leaf senescence by retaining more leaf nitrogen and chlorophyll. Accordingly, it was aimed to study the influence of PGRs and nutrients on the productivity of blackgram.

Materials and Methods

The field experiment was conducted during the kharif season 2002 at Coimbatore. Experiment was conducted in Completely Randomized Block Resign involving CO.6 variety. Seeds were sown in rows 30 cm apart with plant to plant spacing of 10 cm and the population was maintained by thinning. The recommended package of practices were followed. The spraying of 2% DAP (T2), 100 ppm salicylic acid + 2% DAP + 1% KC1 (T3), 100 ppm salicylic acid + DAP 2% + 0.2% Boric acid + 0.5% FeSO₄ + 0.5% ZnSO₄, 0.05% Na₂ MoO4 (T4), 2% urea (T5), urea foliar spray (equivalent N content as in DAP) (T6), humic acid 20 kg soil application + 0.1% humic acid foliar spray (T2), 1% KC1 + humic acid 20 kg/ha as soil application + 0.1% foliar spray (Te), 1% KC1 + humic acid 20 kg/ ha as soil application (To), 0.1 ppm Brassinosteroid (Tto) and control (T1) were done at first flowering (40 DAS) and 15 days after first spray. Nutrients and PGRs were sprayed @



Table 1. Effect of nutrients and PGRs on growth attributes in blackgram

Treatments		Plant height (cm)	4	7.8	LA/plant (cm²)		T	TDMP/plant (g)	3)
	Vegetative	Flowering	Pod filling	Vegetative	Flowering	Pod filling	Vegetative	Flowering	Pod filling
T	17.1	43.1	. 52.5	308.65	594.03	529.10	5.99	7.67	6.82
T.	202	57.1	63.5	508.24	772.69	798.24	7.53	14.05	16.25
'Ľ	21.5	582	61.5	393.05	672.04	721.49	. 734	923	12.94
'n.	17.2	909	0.99	354.49	679.91	726.20 "	99'9	10.31	1131
Ľ	24.4	623	68.0	573.69	924.7	966.50	921	12.01	24.84
, T	18.7	59.4	67.3	405.96	664.7	766.50	626	15.52	15.71
Ţ,	17.7	56.4	62.7	444.53	666.11	890.81	6.79	828	21.22
Ľ	21.4	55.2	62.6	521.01	644.43	896.63	7.61	10.90	16.55
T.	22.1	59.6	65.5	557.52	863.73	899.41	19.9	16.42	23.65
T,	18.4	613	65.8	463.54	655.03	765.25	669	11.71	15.56
Mean	19.55	56.23	63.05	455.82	753.17	836.47	695	11.73	16.95
CD (P=0.05	0	0.1662	SN	229	4.02	4.83	0.0647	0.0967	623

600 lit/ha along with 0.1% Teepol and the control set was sprayed with deionized water and Teepol. Plant height, leaf area and drymatter accumulation was recorded by selecting five random plants from marked rows. The maximum plant height was measured from the base of the stem to the tip of the longest trifoliate leaf. Maximum root length was measured from the base of the stem to the tip of the longest root. Leaf area was measured by using Leaf area meter (LICOR 3000). Drymatter accumulation of various plant parts was estimated by keeping the sample in oven at 80° C for 48 hrs. Total drymatter was quantified by taking the sum of all the plant parts. Growth characteristics namely CGR, NAR, LAD were calculated by adopting the procedure of Watson (1958), Williams (1946), Power et al. (1967) and expressed in g/m2/day, mg/cm2/day and days respectively.

Biochemical estimates viz., soluble protein (Lowry et al., 1951), nitrate reductase activity (Nicholas et al, 1976) and total chlorophyll content (Yoshida et al., 1971) was estimated and the units were expressed as mg/g of fresh weight, μgNO₂/g/ha and mg/g of fresh weight respectively. At harvest pods/plant, 100 seed weight, biological yield and HI were determined from five randomly selected plants. The data were analysed statistically using the 'F test and critical difference (C.D) was calculated (Panse and Sukhatme, 1961).

Results and Discussion

The plant height, leaf area and TDMP increased from vegetative to pod filling stage (Table 1). For plant height, application of 2% urea (T_5) recorded a significant increase (68 cm) over the rest of treatments followed by nutrient mixture with salicylic acid (T_4). The leaf area showed maximum value at pod filling stage when it was sprayed with

Treatment	CGR (CGR (g/m²/day)	NAR (m)	NAR (mg/cm²/day)	LAD	LAD (days)
	35-50 DAS	50-65 DAS	35-50 DAS	50-65 DAS	35-50 DAS	50-65 DAS
ri.	15.7	52	0,210	0.230	25.0	20.5
I,	14.6	8'9	0.240	0.260	33.0	38.5
T,	.681	7.5	0.250	0.340	26.5	27.0
Į.	19.4	8.7	0.280	0.410	23.5	35.0
Ts	24.6	12.8	0.370	0.580	31.0	41.0
T	16.7	6.1	0.240	0.370	36.0	48.0
T,	20.8	92	0.300	0.460	29.0	41.5
H.	21.3	10.8	0.280	0.320	29.5	38.5
Т,	22.5	11.6	0.330	0.550	40.5	48.5
T10	20.5	6.7	0.300	0.540	29.0	35.5
Mean	19.8	8.84	0.300	0.400	30.61	38.47
CD (5%)	0.4852	0.6492	0000	0.011	0.1240	0.1827

chemicals and PGRs. Reduction in leaf area was observed at pod filling stage. Maximum photosynthetic surface was recorded in T5 followed by To. TDMP reflects the accumulated carbohydrate in the plant. Urea spray (T₅) had a profound effect in increasing the TDMP at pod filling stage. Application of PGR increased the TDMP from vegetative to pod filling stage. In the present study, the data revealed an increasing in plant height by urea may be attributed by increasing the N status in the plant system. Thus nitrogen containing source (urea) has increased the height substantially compared to the rest of the nutrients (Kalarani, 1991). The maximum LA at peak flowering contributes to better yielding ability in grain legumes (Thandapani, 1985) is a pre-requisite to maximise the photosynthetic activity. In contrast, in control plants the total LA declined at later period due to the onset of senescence phenomenon (Kalarani, 1991 and Sujatha, 2001). The urea sprayed plant maintained comparatively longer LA at different stages, thus aiding in the supply of photosynthates for the development of pods and grains and also intensification of metabolic activity and efficient utilisation of N (Beninger, 1978). From the result, the reduced TDMP might be due to the reducing photosynthetic activity during senescence period. This would have been brought about by the inability of the plant to take up the nutrient from the soil and also due to the depletion of nutrient from leaves to seed. But application of urea increased the TDMP by reducing the leaf senescence (Syverud et al. 1980 and Kannan, 1986).

Application of PGR and nutrients increased the growth characters viz., CGR, NAR and LAD (Table 2) at

Table 3. Effect of nutrients and PGRs on biochemical attributes in blackgram

Treatments	Total chlorophyll (mg/g)		Soluble protein (mg/g)		NRase activity (ug NO ₂ N/g/ha)	
	Vegetative	Flowering	Vegetative	Flowering	Vegetative	Flowering
T ₁	0.4783	2.0687	08.00	10.47	24.33	29.00
T ₂	0.6203	2.1031	10.20	13.30	26.60	30.31
T_3	0.8869	2.4236	10.64	13.70	31.00	38.30
T ₄	0.9449	2.4826	11.29	14.10	40.34	47.33
T ₅	1.8884	2.8623	12.65	15.40	49.62	56.60
T ₆	0.8696	2.3839	11.26	13.10	32.02	42.00 -
T ₇	1.0460	2.5165	11.67	13.20	44.33	50.30
T ₈	0.8289	2.3435	11.60	13.70	32.51	40.63
T ₉	1.0928	2.6850	12.02	15.00	46.31	53.62
T ₁₀	0.9304	2.4788	10.73	12.10	46.61	51.30
Mean	0.8773	2.4517	11.005	13.407	37.29	44.50
CD(P=0.05)	0.0045	0.0057	0.0148	0.0160	2.164	2.396

Table 4. Influence of nutrients and PGRs on yield and yield attributes in blackgram

Treat- ments	Number of pods/ plant	Number of clusters/ plant	100 seed weight (g)	Grain yield/ plant(g)	Yield (kg/ha)	н	- (+
T ₁	21.31	8.44	3.92	3.74	602.20	0.20	
T ₂	22.43	10.97	3.97	4.46	690.50	0.27	
T ₃	26.52	11.86	4.20	4.53	808.80	0.25	
T_4	35.60	12.53	4.26	4.65	849.30	0.30	
T ₅	44.72	15.84	4.48	5.10	955.20	0.32	
T ₆	24.36	11.76	4.11	45.12	806.00	0.28	
T ₇	40.93	13.89	4.28	4.77	872.50	0.31	
T ₈	26.14	11.59	4.02	4.92	752.20	0.27	
T ₉	43.81	14.91	4.44	5.02	926.20	0.32	
T ₁₀	30.55	12.08	4.22	4.84	816.30	0.29	
Mean	31.64	12.39	4.23	4.62	813.58	0.29	
CD(P=0.05)	0.867	0.15	0.03	0.026	2.52	0.009	

all growth stages. Among the treatments, T5 recorded a maximum CGR value of 24.6 g/m²/day followed by T₀ (22.5 g/m²/day) at 30 - 50 DAS. Thereafter CGR showed a declining trend. In T5 and To CGR decreased to a tune of 50% from the critical (35 -50 DAS) value. More LAD was obtained in T_q (48.0). It increased from 35-50 DAS to 50 - 65 DAS. The decreasing in LAD in control at lateral period of reproductive phase may be due to decreased number of leaves and leaf surface. All other treatments increased the LAD which is an essential factor for increased the photosynthesis during grain filling period. The increased LAD may be due to increased LAI of urea treated plants. Maximum NAR was recorded at urea treatment (T₅), higher NAR at 50 - 65 DAS may be due to rapid vegetative growth, since the leaf nitrogen is related with NAR during grain filling period which enables the plant to have higher NAR (Calmes et al., 1989, Kalarani, 1991 and Sujatha, 2001). The increase in CGR is due to chemical spray may be attributed to increased LAD. As CGR is a linear function of intercepted irradiation (Shibles and Webber, 1965; 1966) and maintenance of higher LAD has positive effect on higher drymatter production ultimately resulting in higher yield (Gopal Singh and Jainarain Singh, 1982).

The total chlorophyll, soluble protein and NR activity increased from vegetative stage to flowering stage. Among all the treatments, urea had the profound effect in improving the total chlorophyll content, soluble protein content and NR activity followed by the next best treatment T₉ (Table 3). The increased chlorophyll content in urea sprayed plants may be due to the fact that nitrogen is a constituent of chlorophyll molecule which is expected during rapid grain filling leaves (Mitra et al., 1987 and Sujatha, 2001). Martignone et al. (1987) reported that soluble proteins are the first nitrogenous compound lost during pod

filling and spraying of urea could increase the soluble protein content. This is in conformation with the finding of Manian et al. (1987). The NR activity is associated with availability of carbohydrate (Luis and Autonio, 1983). The increased activity is due to urea spray may be attributed by increased enzyme synthesis and high NAR (Mirhatum and Hume, 1976; Akhtar et al. 1991).

Foliar spray of 2% urea had profound effect in improving the yield and yield attributes characters viz., number of pods per plant, number of clusters per plant, 100 seed weight, grain yield per plant and yield per hectare (Table 4). The number of pods and clusters per plant were increased due to the application of urea to a tune of 105 per cent and 87 per cent over control followed by T₉. Urea application had the significant yield increase of 58.6 per cent when compared to control followed by the treatment T₉ (53.8%). However, both the treatments (T₅ and T₉) which are on par with each other for harvest index.

As pod number is considered to be the major yield determinant in pulses, foliar feeding of N through urea source was able to increase the pod number in this experiment. This observation is in confirmation with the report of Brevedan et al. (1978). The data illustrate the importance of the N nutrition of blackgram plant during reproductive stage. Lathwell and Evans (1951) have reported similar results. The data on 100 seed weight showed that control plants recorded lesser seed weight. The reduction in seed weight may be due to the change in the flux of cytokinin and mineral nutrients from the roots normally declines during pod filling (Nooden, 1984 and Nooden et al. 1989) and also by complete deprivation of cytokinin and minerals from this source. Improvement of accumulation of biomass on seed due to other selected chemicals may be attributed due to increased source activity by delaying the senescence during grain filling period. The higher yield noticed in the effective treatments may probably due to longer retention of the effective photoassimilatory surface. Thorne (1973) and Sekhan et al. (1987) has suggested that delay in leaf senescence may be achieved agronomically through application of nitrogen fertilizer at flowering. HI and yield are directly correlated with yield in pulses (Kalarani, 1991). In the present study also high yielders had high partitioning efficiency.

Conclusion

The present investigation reveals that the foliar spray of plant growth regulators and chemicals were able to influence the physiological, biochemical and yield components of the crop. In particular, foliar spray of 2% urea was found to be the most effective treatment followed by KCI 1% along with soil application of humic acid @20 kg/ha. Though the selected chemicals were able to increase the yield, 2% urea was found to be the best than others due to efficiency in increasing the yield.

References

- Akhtar, ML, M.M.Samiulkh, R.K.Afridi and F.Khan. (1991). Correlation of nitrate reductase activity with yield and protein content of lentil. Comparative Physiology and Ecology, 14: 103-107.
- Amal D. A. (2001). Effect of humic acid treatments on growth and development of groundnut under different fertilizer levels. M.Sc.(Ag.) Thesis, Tamil Nadu Agricultural University, Coimbatore.
- Beninger, H. (1978). The role of potassium in yield formation. Potash Review Subject 16, Suite 78, Int., Potash Inst Berne (Switzerland), pp.3.
- Brevadan, R. E., D. B. Egli and J. E. Leggert. (1978). Influence of N nutrition of flower and pod abortion and yield of soybeans. Agron. J., 70: 81-84.

- Calmes, J., M.Bensari, G.Viala and N.Gelfi. 1989. Soybean leaf assimilates and their utilization in seed filling and the effects of nitrogen application. Field. Crop Abst. (1990), 43: 45.
- Gopal Singh, G. and Jainarain Singh. (1982). Effect of seasonal changes on growth parameters of greengram. Indian J. Plant Physiol., 25: 382-389.
- Kalarani, M.K. (1991). Senescence regulation in soybean. M.Sc.(Ag.) Thesis, Tamil Nadu Agricultural University, Coimbatore.
- Kannan, S. (1986). Foliar absorption and transport of inorganic nutrients. CRC critical reviews. (Plant Sciences CRC Press, Inc., USA, in press).
- Lathwell, D.J. and C.E. Euans. (1951). N uptake from solution by soybeans at successive stages of growth. Agron. J., 43: 264-270.
- Luis, G. S. and C. N. M. Autonio. (1983). Changes in nitrate reductase activity during development of soybean leaf. J. Plant Physiol., 112: 113-121.
- Lowry, O.K., N.T. Rose Brought, L.A. Farr and R.J. Randall. (1951). Protein measurement with folin phenol reagent. J. Biol. Chem., 193: 265-275.
- Manian, K., N.Natarajaratnam, P.Ramasamy and K.Mohanasundaram. (1987). Senescence and its role in soybean productivity. J. Agron. Crop Sci., 159: 202-205.
- Martignone, R.A., J.J. Guiamet and F.Nakayama. (1987). Nitrogen partitioning and leaf senescence in soybean as related to nitrogen supply. Fid. Crops Res., 17: 17-24.
- Mirhatum and D.J.Hume. (1976). Relations between nitrate reductase activity and nitrogen accumulation in soybean. Can. J. Plant Sci., 56: 377-384.