https://doi.org/10.29321/MAJ.10.A00119 ffect of sources, concentration and frequencies of B application on he petiole nutrient content in graphs (Vitis vinifera) cv. Muscat.

). JANAKI*, V. VELU AND P. SAVITHIRI

Department of crop Management, ADAC & RI, Trichy Dept. of Soil Science and Agrl. Chemistry, Tamil Nadu Agrl. University, Coimbatore - 641 003.

Abstract: A field expetiment was conducted at Mathampatti to study the effect of B spray on the petiole nutrient content of the grapes at full bloom stage. The study revealed that N, P, K content of the potiole increased with B application while a depressive effect was noticed on Ca content. Micronutrients like Cu, B, Zn and Fe content were also found to increase with B application.

Key words: Agribor, Boricacid, Concentration of spray, Frequency of spray, Petiole nutrient

ntroduction

Grape is subtropical fruit and its cultivation s fast expanding. It is taken up on wide range of soils in different agrolimatic situations of ndia. In Tamilnadu it is grown in selected listricts of Theni, Dindugal, Madurai and Coimbatore. Generally five crops are grown in 2 years. Due to continuous fruting habit and faster growth and heavy yield, it require nutrient supply throughout its growth period. The yields are found to be erratic due to depletion of secondary and micronutrients (Tandon, 1993). Leaf analysis serves as an ideal indicator of the nutritional status of the plant at any time during growing season (Bhargava and Chadha, 1988). Petiole samples collected at flowering stage will helps to monitor nutrition for quality rather than yield (Bhargava and Summer, 1987). However, the information regarding the petiole nutrient status of grapes is lacking under local conditions. Hence, the present study was carried out to study the impact of boron spray on the petiole nutrient content at full bloom stage.

Materials and Methods

The present study was carried out in the farmers holding at Mathampatti during the year 2000-2001 on a fully grown 31/2 year old grapevine. The experiment was carried in

a Typic Haplustalf, sandy clay loam in texture with a pH 7.7, available N 210 kg ha-1 Olsen P 84.6 kg ha-1 and Hotwater soluble B-0.42 mg kg-1. The trial was laid out in a split plot design with 2 sources of boron viz. agribor and boric acid comprising of 3 concentrations of spray viz. water spray, 0.1% spray, 0.2% and 0.3% spray with three frequencies viz. one, two and three times at three critical stages, viz. bud differentiation stage, full bloom and 15 days after full bloom. The recommended dose of NPK @ 260:160:600g per vine was applied for all the plots and all other cultural operations were carried out uniformly. The petiole samples opposite to flower cluster were collected from each plot at the full bloom stage (Bhargava and Chadha, 1988). The petiole samples were washed with 0.1N HCl followed by double distilled water, dried and powered in a stainless steel whileymill and the samples were digested in diacid extract and the extract was analysed for nitrogen (Humphries, 1956). Triple acid extract of the sample was used for the estimation of phosphorus (Jackson, 1953); K (flamephotometry); B (Page et al. 1983); Ca (Versenate method); microinutrients (Lindsay and Norwell, 1978).

Results and Discussion

Petiole nitragen, phospharus and potassium

The petiole nutrient content of Muscat grapes sampled at full bloom stage indicated

Table 1. Effect of sources, concentrations and frequencies of B spray on the petiole N, P and K content of grapes.

(Mean of 3 replications)

i. Nitorgen content (%)

Concentration		Frequen	су	- 30 - 41	Sour	Mean	
	F _i	F ₂	F ₃		Sı	S ₂	Se - 19 15
C _o - Control	1.63	1.67	1.66	Ħ	1.65	1.66	1.65
C 0.1%	1.64	1.68	1.77		1.72	1.68	1.70
C ₂ - 0.2%	1.75	1.78	1.75		1.79	1.75	1.77
C, - 0.3%	1.75	1.76	1.77	-	1.79	1.74	1.76
Mean	1.69	1.72	1.74	4)	1.74	1.71	1.72
	s	C	F	SxC	SxF	CxF	SxCxF
SEd	0.003	0.013	0.007	0.03	0.002	0.019	0.046
CD (P=0.05%)	NS	0.02	NS	NS	NS	0.04	. NS

ii. Phosphorus content (%)

Concentration	* **	Frequenc	y .		Sour	Mean	
	F,	F ₂	F,		S,	S ₂	ti.//
Co - Control	0.43	0.44	0.45		0.43	0.46	0.44
C, - 0.1%	0.36	.0.34	0.43		0.37	0.44	0.40
C ₂ - 0.2%	0.40	0.35	0.40		0.40	0.43	0.41
C, - 0.3%	0.44	0.44	0.45		0.45	0.43	0.44
Mean	0.41	0.39	0.43		0.41	0.44	0.42
	S	C	F	SxC	···SxF	CxF	SxCxF
SEd	0.001	0.01	0.06	0.02	0.01	0.02	0.03
CD (P=0.05%)	NS	0.02	NS	NS	NS	NS	NS .

iii. Potassium content (%)

Concentration	36	Frequenc	y		Sour	Mean	
	F ₁	F ₂	į.	F,	S	S ₂	
C _o - Control	2.30	2.32		2.26	2.35	2.24	2.37
C, - 0.1%	2.33	2:49		2.58	2.49	2.45	2.48
C, - 0.2%	2.53	2.78	p 1	2.36	2.62	2.49	2.55
C, - 0.3%	2.33	2.33		2.47	2.42	2.34	2:38
Mean .	2.37	2.48		2.42	2.47	2.38	2.42
e. Free	S	C	F	SxC	SxF	CxF	SxCxF
SEd	0.03	0.03	0.02	0.04	0.05	0.07	0.07
CD (P=0.05%)	0.13	0.06	0.05	NS	0.10	NS	NS:

Table 2. Effect of sources, concentrations, frequencies of B spray on the petiole Ca (%), B and Zn (mg kg⁻¹) of grapes.

(Mean of 3 replications)

i. Calcium content (mg kg-1)

Concentration		Frequency			Source		
-	F,	F ₂	F,	S,	S ₂	Mean	
C ₀ - Control C ₁ - 0.1%	1.84	1.74	1.82	1.78	1.82	1.80	
C ₂ - 0.2%	1.82 1.77	1.82	1.71	1.79	1.80	1.79	
C, - 0.3%	1.65	1.81	1.70	1.76	1.76	1.76	
Mean	1.77	1.74	1.65	1.68	1.67	1.67	
•		1.78	1.72	1.76	1.76	1.76	
SEd	S	C	F Sx	U.J. 33,779.	CxF	SxCxF	
CD (P=0.05%)	0.03 NS	0.02	0.02 0.0		0.05	0.05	
CD (1-0.05%)	142	0.03	NS N	S 0.12	·NS	NS	

ii..Boron content (mg kg-1)

Concentration		Frequen	су		Sour	Mean	
	F ₁	F ₂		F,	S,	S ₂	-
Co - Control	162	171	247	193	176	174	175
C, - 0.1%	214	217	1 2	219	217	217	217
C ₂ - 0.2%	221	219		221	222	218	220
C, - 0.3%	220	224		226	225	222	222
Mean	204	208		215	210	207	208
	S	C.	F	SxC	SxF	CxF	SxCxF
SEd	1.6	2.6	2.7	3.73	5.41	3.82	9.3
CD (P=0.05%)	NS	5.3	NS	NS	NS	NS	18.6

iii. Zinc content (mg kg-1)

Concentration		Frequency				ce	Mean
	F,	F,	F,		S,	S ₂	
C _o - Control	36.3	36.8	37	5	36.1	34.5	35.3
C 0.1%	37.1	37.7	38.	4	37.8	37.6	37.7
C, - 0.2%	38.8	39.8	39.	7	39.1	39.7	39.4
C, - 0.3%	39.2	38.3	37.	3	38.6	38.1	38.3
Mean .	37.8	38.1	38.2	2	37.9	37.5	37.7
4.	s	С	F	SxC	SxF	CxF	SxCxF
SEd	0.39	0.40	0.35	0.65	0.71	0.87	1.23
CD (P=0.05%)	NS	0.9	NS	NS	NS	NS	NS

Table 3. Effect of sources, concentrations, frequencies of B spray on the petiole Cu and Fe and Mn (mg kg⁻¹) of grapes.

(Mean of 3 replications)

i. Copper content (mg kg-1)

Concentration		Frequency				Source		
	F,	F ₂	F,		S,	S ₂		
C _o - Control	110	116	114		113	114	113	
C, - 0.1%	116	119	122		119	120	119	
C 0.2%	121	115	114	*	116	120	118	
C, - 0.3%	113	117	118		117	115	116	
Mean	115	117	117		116	117	116	
	S	C	F	SxC	SxF	CxF	SxCxF	
SEd	1.28	1.15	1.05	2.00	2.10	1.91	2.98	
CD (P=0.05%)	NS	2.3	NS .	NS	NS	NS-	NS	

ii. Iron content (mg kg-1)

Concentration	4	Frequency			Sour	ce .	Mean
	F ₁	F ₂	F,		S	S ₂	
Co - Control	197	.192	190		198	194	196
C, - 0.1%	200	197	202		201	199	200
C ₂ - 0.2%	206	203	204		205	205	205
C, - 0.3%	207	193	183		191	193	.192
Mean	202	196	195	٠	199	198	198
19	S	C .	F	SxC	SxF	CxF	SxCxF
SEd	0.94	2.80	2.00	3.83	4.69	4.06	6.60
CD (P=0.05%)	NS	5.7	NS -	NS	NS	NS	NS

iii. Manganese content (mg kg-1)

Concentration		Frequenc	y	4	Source		
	F _t	F ₂	. F ₃		S,	S ₂	- h _
Co - Control	54.4	54.7	54.8	1 14	54.8	54.5	54.6
C, - 0.1%	55.0	54.9	55.9		55.6	55.0	.55.3
C, - 0.2%	55.8	56.7	56.4		56.4	56.2	56.3
C, - 0.3%	55.8	55.6	56.3		55.3	56.3	55.8
Mean	55.2	55.5	55.9		55.5	55.6	55.5
	S	C	F	SxC	SxF	CxF	SxCxF
SEd	0.64	0.82	0.56	1.42	1.12	0.97	1.59
CD (P=0.05%)	NS	NS	NS	NS	NS	NS	NS

that foliar spray of boron increased the petiole N content. The mean values ranged from 1.65 to 1.77 per cent. Among the concentrations of spray, the effect of 0.2 and 0.3 per cent were comparable. In concentration x Frequency interaction, 0.1 per cent spraying thrice was effective in increasing the petiole N content (Table 1). Increased frequency increased the petiole N content which might be due to the positive interaction of N and B in plant system as reported by Yadav and Manchanda (1979). The petiole P content was increased at 0.3 % spray followed by 0.2% spray. The highest P content was recorded at 0.3% (0.44 %) and it was similar to the findings as reported by Prabha (1995). The agribor recorded higher K content. Among the concentrations, 0.2% recorded higher K in the petiole as well as wo times spray registered the highest K content (2.48%). In concentration x frequency interaction, 3.2, per cent at one or two times spray was peneficial. This might be due to the positive effect of B on K nutrition

Petiole calcium, boron and zinc

The results are given in Table 2. Petiole Ca decreased with B application. The petiloe Ca decreased as the concentration increased beyond 0.1% and result could be seen in both the sources. Similar reduction in the content was noticed by Marhorkar and Patel (1987).

Petiole B content was markedly increased with increased concentration. The highest B content was observed at 0.3 per cent (222 mg kg-1) while lowest in control (175 mg kg-1) (Table 2). Increase in the frequency of spray brought out relative increase in the B content in the petliole and this might be due to effective translocation of boron to petiole due to direct supply of B to the foliar tissue through foliar spray as reported by Shorrocks (1997). Petiole Zn was increased due to B treatments. Increase in concentration of spray increased the Zn concentration in the petiole. Application of 0.2 per cent (39.4 mg kg⁻¹) spray registered increasing petiole Zn content whereas further increase result in decline in petiole Zn content.

Petiole Copper, Iron and Manganese:

Copper content in the petiole was the highest in 0.1 per cent spray (199 mg kg⁻¹) while control registered 113 mg kg⁻¹ of Cu. This was due to the positive correlation between B and Cu as reported by Sabanayagam (1996). Increase in concentration of B spray increased the Fe content at 0.2 per cent (205 mg kg⁻¹) while a further increase in concentration caused a reduction in petiole Fe (Table 3.) This corroborates with the findings of Singh et al. (1990). Manganse content was not much influenced by the B treatments.

Conclusion

The results of present study emphasized that application of foliar of boron significantly increased the N, P and K content in the petiole while the Ca content decreased with increase in boron spray due to negative interaction of Ca with B. As regards to micronutrients, Cu, Mn increased while Fe content decreased.

References

Bhargava, B.S. and Chadha, K.L. (1988). Developing leaf nutrient guide in fruits and plantation crops. Fert. News, 33: 12-29

Bhargava, B.S. and Summer, M.E. (1987). Proposal for sampling grape (vitis vinifera) petioles for nutritional diagnosis. Comman. Soil Sci. Plant Anal., 18: 581-591.

Humphries, EC. (1956). Mineral components and ash analysis: modern method of plant analysis. Springler verlag Berlin, 1: 418 - 502

Jackson, M.L. (1973). Soil Chemical analysis. Prentice Hall Private limited, New Delhi.pp. 498

Lindsay, W.L. and W.A. Norvell. (1978). Development of DTPA soil test for inc, Iron, manganese and copper. Soil.Sci.soc. Amer. J., 42: 421-428

Marhorkar, V.K. and Patel, V.K. (1987). Effects of boron on growth of the cultivars of grape. J. Maharashtra Agric. Univ., 12: 394-395.

- Page, A.L. R.H. Miller and D.R. Keeney. (1982). Method of soil anaysis. Part 2 Sr. No.9 Am. Soc. Agron., Inc., Madison, U.S.A
- Prabha, K. (1995). Studies of boron nutrition of Tomato in calcarcous soils. M.Sc.(Ag.) thesis submitted to Tamil Nadu Agricultural University, Coimbatore
- Sabanayagam, V. (1996). Appraisal of available boron status of Tamil Nadu soils and its fertilization to groundnut - sunflower cropping system in Entisols Ph.D. Thesis submitted to TNAU, Coimbatore-3.
- Shorrocks. V.M. (1997). The occurrence and correction of boron deficiency In: Boron in soils and plants. (Ed). R.W.Bell, B.Bell and P.H. Brown. Klumer Academic Publishers, Netherland. Pp.121-148.

- Singh, B., Revathy, P. and Prasad, R (1990). Effect of microcnutrient on the physical and biochemical parameters of kazi lime. Prog. Hort, 22: 316-219
- Tandon, H.L.S. (1993). Method of analysis of soils, plant, water fertilizer. Fertilizers Decvelopment and Consultation Organization, New Delhi. India.
- Yadav, O.P., Manchanda, H.R. (1979). Boron tolerance studies in gram and wheat grown in Sermozem sandy soil. J. Indian Soc. Soil Sci., 27: 173-180.
- (Received: July 2003; Revised; May 2004)