## GRANULATION OF VA-MYCORRHIZAL INOCULUM

S. SUJATHA LILLY and P. SANTHANAKRISHNAN Department of Agricultural Microbiology TNAU, Coimbatore - 641 003.

#### SUMMARY

An attempt was made to develop a granular VA - mycorrhizal inoculum for easy application to polythene bags as well as nursery raised plants. The clay was used as a substrate for preparation of granules. The granules were prepared with VA-mycorrhizal fungi individually and in combination with Azospirillum and Phosphobacteria. The shelf life of granular formulation of VAM fungi with Azospirillum and Phosphobacteria. The shelf life of granular formulation of VAM fungi with Azospirillum and Phosphobacteria was tested and found that there is a possibility of storing the granules upto 60 days. The response to inoculation of granular formulation prepared with all the three inoculants was superior than two inoculants or uninoculated control.

KEY WORDS: Granular inoculum, VAM fungi, Azorpirillum.
Phosphobacteria

Mycorrhizal plants exhibit greater of phosphorus and other micronutrients particularly when those nutrient are sparingly soluble in soil (Abbott and Robson, 1984) which inturn increase the growth and yield of different crop plants. Azospirillum, an associative nitrogen fixing bacterium and phosphate solubilising bacterial are widely used in increasing the growth of crop plants. However these inoculants are used singly. Hall (1979) developed a VAM fungal inoculum which include a mixture of soil, roots and spores adhered to seeds with adhesives Sweet potato entrapped in responded to early and rapid colonization by effective strains of VAM fungi over a practical range of P fertility (Hung et al., 1994). Strullu and Plenchette (1990, 1991) encapsulated the active intraradical form of Glomus intraradices in calcium alginate beads. But the disadvantage of this method is that hydrogels are expensive.

VA-mycorrhizal inoculum can come in a range of forms such as soil, spore preparation, dried mycelium, infected roots or more sophisticated products such as alginate beads with incorporated mycelial homogenates. The soil based inocula is too heavy for extensive use in agriculture. For efficient handling, vesicular-arbuscular mycorrhizal fungi may be processed into small and uniform inocula, however processing can reduce the inoculum density. One of the main problems in inoculant technology is the survival of microorganisms during storage. There is a need to develop a quality inoculum and formulation having VA-mycorrhizal fungi and other beneficial

microorganisms for easy application to nursery raised plants. Hence a study was conducted to develop a granular formulation which consists of VAM along with Azospirillum and Phosphobacteria and tested the response in maize plant due to inoculation of granular formulation.

### MATERIALS AND METHODS

# (a) Procedure for preparation of granular VAM fungi inoculants

VA- mycorrhizal bulk inoculum (20 g) consisting of infected root, spores and hyphae was mixed with two gram of sterilized clay soil. These materials are mixed well and granules were prepared of pellet size I am size 96 hour old Azospirillum broth (5 x 10° cells/ml) / 72 hr old Phosphobacteria broth (2 x 107 cells/ml) were used instead of sterile distilled water for preparing inoculants which consist of Azospirillum or Phosphobacteria. For granules which consist of both inoculants equal quantity of mixer is used for preparing the granules.

(b) The effective response of granular VAM fungi inoculants incorporated with Azospirillum and Phosphobacteria were tested by using maize as a host plant.

The treatment details are as follows:

- 1. Glomus fasciculatum
- 2. Glomus fasciculatum + Azospirillum brasilense
- 3. Glomus fasciculatum + Phosphobacteria (Ps-1)

Table 1. Survival of Azospirillum population in VAM granular inoculum incorporated with Azospirillum and Phosphobacteria

F	-	Azospirii	lum population x	10th cells g-1*	*
Treatment		ř.			
	1	15	30	45	60
Glomus fasciculatum				\$ <b>#</b> /	
Glomus fasciculatum + Azospirillum	5.4	4.3	2.1	1.5	0.84
Glomus fasciculatum + Phosphobacteria	<i>i</i> ≠ #.	Ξ	-	÷/-	.1*:
Glomus fasciculatum + Azospirillum +	4.3	3.5	1.9	1.1	0.64
Phosphobacteria Control	<u>-1</u>	<u>.</u>	2		¹. <b>≱</b> .

<sup>\*</sup> Average of five replications

### 5. Uninoculated control

The treatments were replicated with 10 plants per replication. Plant samples were taken from each treatment on 45th day after sowing and the shoot length, root length, plant dry weight were recorded. The VA-mycorrhizal spore count was

determined by wet sieving and decanting technique of (Gerdemann and Nicolson, 1963) and the percentage mycorrhizal colonization by staining roots with tryphan blue (Philips and Hayman, 1970) The dry weight of extramatrical hyphae was estimated using the membrane filter technique of Abbott et al. (1984). Population of Azospirillum in the granules and rhizosphere soil was estimated by adopting MPN technique (Okon

Table 2. Survival of Phosphobacteria population in VAM granular inoculum incorporated with Azospirillum and Phosphobacteria

		Phospho	bacteria population	n x 10° cells g-1°	
Treatment			Days after storage		
	i İ	15	30	45	60
Glomus fasciculatum					
Glomus fasciculatum + Azospirillum	-	2.	i( <b>.</b> c		+
Glomus fasciculatum + Phosphobacterie	10.5	8.5	7.5	4.0	2.5
Glomus fasciculatum + Azospirillum +	7.5	6.5	6.0	3.5	2.0
Phosphobacteria Control	-	٠.,			

<sup>\*</sup> Average of five replications

Glomus fasciculatum + Azospirillum brasilense + Phosphobacteria

 et al. 1976) and Phosphobacteria by enrichment
 culture technique described by Katznelson and Bose (1959)

### RESULTS AND DISCUSSION-

Initial moisture content and water holding capacity on the day of preparation of granules was 12.1 percent and 54.69 percent which was reduced to 11,94 percent when the granules were stored for 60 days. Population of Azospirillum and Phosphobacteria survived well in the granular formulation even after a storage period of 60 days (Table 1 and 2). The efficiency of granular inoculum was tested in maize and the results revealed that by using formulation prepared with Glomus fasciculatum, Azospirillum and Phosphobacteria showed increased growth, plant dry weight (Table 3), VAM colonization (Table 4), and dry weight of extramatrical hyphae when compared to twin inoculants or uninoculated control at 45 days after sowing. In the present study, VAM fungi survived well in the dried granules and infected the maize roots even when 60 day old granules were used.

The entrapment of microbial cells could enhance the shelf life of the organisms and there is a possibility for combining two or more organisms. VA-mycorrhizal inoculation significantly improved the growth and uptake of Phosphorus. Similar result was obtained by Hall (1979) who reported that white clover plants inoculated with infested soil pellet containing VAM fungi recorded maximum shoot length. VAM colonization and uptake of phosphorus when compared with uninoculated control. Better survival of Azospirillum in alginate formulations was reported by Roughley (1968) and Fages (1990). The present investigation clearly indicates that granules which consists of Glomus fasciculatum, Azospirillum and Phosphobacteria increased the growth, nutrient uptake and VAmycorrhizal colonization than pellets having combination of two inoculants. The improved growth can be attributed to cumulative, greater nutrient uptake and production of growth promoting substances by Azospirillum and Phosphobacteria.

Table 3. Influence of VAM granular formulation incorporated with Azospirillum and Phosphobacteria on the dry weight of host plant (maize)

Treatment —	Dry weight (g/plant)*  Days after storage						
	Glomus fasciculatum	0.696	0.689	0.635	0.617	0.625	0.653
Glomus fasciculatum + Azospirillum	0.731	0.732	0.725	0.723	0.719	0.726	
Glomus fasciculatum + Phosphobacteria	0.831	0.836	0.828	0.819	0.821	0.828	
Glomus fasciculatum + Azospirillum +	1.272	1.099	1.146	1.169	1.165	1.170	
Phosphobacteria							
Uninoculated control	0.530	0.527	0.519	0.519	0.560	0.531	
Меап	0.812	0.777	0.771	0.769	0.778	0.781	

<sup>. - 45</sup> days after Sowing

	SEd	CD
Treatment	0.017	0.033
Stages	0.017	- 0.033
Treatment x stages	0.037	0.073

Table 4. Influence of VAM granular formulation incorporated with Azospirillum and Phosphobacteria on per cent root colonization of Glomus fasciculatum in maize

	*5		Per cent root	colonization*			
Treatment	Days after storage						
	1	. 15	30	45	60	Mear	
Glomus fasciculatum	.34	32	32	29	27	31	
Glomus fasciculatum + Azospirillum	35	33	34	27	26	31	
Glomus fasciculatum + Phosphobacteria -	- 38	35	34	28	27	32	
Glomus fasciculatum + Azospirillum +	39	37	35	28	28	33	
Phosphobacteria							
uninoculated control	14	12	12	13	15	13	
Mean	32	30	30	25	25	28	

<sup>. - 45</sup> days after sowing

-	SEd	CD
Treatment	0.60	1.18
Stages	0.60	1.18
Treatment x stages	1.32	2.64

### REFERENCES

- ABBOTT, L.K. and ROBSON, A.D.(1984). The effect of mycorrhizae on plant growth. In: 'VAmycorrhizae'(Eds. Powell, C.L. and Bagyaraj, D.J.) pp 113-130 CRC Press, Boca Raton.
- ABBOTT, L.K., ROBSON, A.D. and DE BOER, G. (1984).

  The effect of Phosphorus on the formation of hyphae in soil by the vesicular-arbuscular mycorrhizal fungus.

  Glomus fasciculatum New Phytol., 97: 437 444.
- FAGES, J. (1990). An optimized process for manufacturing an Azaspirillum inoculant for crops. Appl. Microbiol. Biotechnol., 32: 473.
- GERDEMANN, J.W. and NICOLSON, T.H. (1963). Spores of mycorrhizal endogone species extracted from soil by wet sieving and decanting Brit. Mycol. Suc., 46: 234 - 244.
- HALL I.R. (1979). Soil pellets to introduce vesiculararhuscular mycorthizal fungi into soils Soil Biol. Biochem., 11: 85 - 86.
- HUNG, L.L., O'KEEFE, D.M. and SYLVIA, D.M. (1994). Use of hydrogels as a sticking agent and carrier for vesicular arbuscular mycorhizal fungi. Mycol. Res., 58: 54-58.

- KATZNELSON, H and BOSE, B. (1959). Metabolic activity and phosphate dissolving ability of bacterial isolates from wheat roots rhizosphere and non-rhizosphere soil. Can. J. Microbiol., 5: 79-85.
- OKON, Y., ALBRECHT, S.L. and BURRIS, R.H. (1976). Carbon and Ammonia metabolism of Spirillum lipoferum. J. Bacteriol., 128: 592-597.
- PHILLIPS, J.M. and HAYMAN, D.S. (1970) Improved process for clearing and staining parasite for vesiculararbuscular mycorrhizal Jungi, Trans. Brit, Mycol. Soc., 55: 158-166.
- ROUGHLEY, R.J. (1968), some factors influencing the growth and survival of the root nodule bacteria in peat culture. J. Appl. Bacteriol., 31: 259-265.
- STRULLU, D.G. and PLENCHETTE, C. (1990). Encapsulation de la forme intracinaire de Glomus dans l'alginate et utilization des capsules comme, inoculu, C.R. Acad., 310 : 447-452.
- STRULLU, D.G. and PLENCHETTF, C. (1991), The entrapment of Glomus sp. in algunate beads and their use as root inoculum. Mycol. Res., 95: 1191-1196

(Received - December 1997 Revised : August 1998)