MOODY, K. (1982). Weed control in dryland crops after rice. In: Report of a Workshop on Cropping Systems Research in Asia.

MOORTHY, B.T.S. and MANNA, C.B. (1982). Weed control in transplanted rice by herbicides in dry season. Abstr. Ann.

Madras Agric. J., 82(5): 387-390 May, 1995 https://doi.org/10.29321/MAJ.10.A01215 Conf., ISWS 13 pp.

RAO, V.S. (1983). Principles of Weed Science. Oxford and IBH publishing Co., New Delhi.

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PHENOTYPIC STABILITY OF GRAIN YIELD AND ITS COMPONENTS IN CHICKPEA

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ABSTRACT

Twenty chickpea genotypes were grown in six environments to study the phenotypic stability of grain yield and its components. It was found that the linear component of G x E interaction was more important for yield and other characters. Four genotypes i.e., SGM 84-104, IH 83-6, SG 2 and SGM 84-117 were found to have average response and high stability and high mean for grain yield. However, SGM 84-112 the highest yielding genotype was highly unstable. There was positive and significant correlation between the mean of the genotypes and the responsiveness for number of pods/plant, 100-grain weight and single plant yield which indicated that the genotypes with high mean were, in general, better responsive to favourable environments. There was lack of general association between stability of yield and its components which calls for cautious selection of genotypes based on yield alone.

KEY WORDS: Cicer arietinum, Chickpea, Phenotypic Stability, Genotype x Environment Interaction.

Chickpea (Cicer arietinum L.) is one of the most important pulse crops of India. Development and adaptation of high yielding varieties appear to be the most important step for increasing production. Although number of improved varieties of gram have been evolved, the yield of these varieties is not stable over environments which is one of the reasons for their poor adaptation. Thus stability is one of the desirable properties of a genotype sought for in a variety. Though the information on genotype x environment (G x E) interaction has been adequately worked out in zereal crops, the relative basic information on hickpea is limited. Therefore, the present nvestigation was planned to collect the information bout stability in chicknea.

MATERIALS AND METHODS

The experimental materials consisted of 20 genotypes of chickpea including 3 varieties viz., BR 77, C 235 and SG 2 and 17 advance generation ines. In all, six environments were created by rowing them in two dates of sowing at two different locations i.e., Bhagalpur and Muzaffarpur luring 1985-86 and 1986-87. The first sowing was sone in the first week of November and second in

the first week of December each year. The entries were sown in a randomised complete block design with three replications in 4m long rows, spaced 30 cm apart, with plant to plant distance of 10 cm. Recommended agronomical practices were followed throughout the crop season. Observations were recorded on five randomly selected plants in each entry on the number of pods/plant, 100 grain weight (g), single plant yield (g) and grain yield (q/ha). Grain yield was calculated from plot yield. The stability analysis was performed according to the model suggested by Eberhart and Russell (1966).

RESULTS AND DISCUSSIONS

lighly significant variances due to genotypes revealed the presence of considerable genetic variability among the genotypes for all the characters studied (Table 1). Highly significant mean squares due to environments and genotype x environment interactions suggested the presence of considerable interactions of the genotypes with the environmental conditions. Highly significant variances due to environment (linear) indicated that creations of the environments by manipulating dates of sowing, locations and years was effective.

Table 1. Analysis of variance for grain yield and its components in chickpea

P	df	M.S.					
Sources		Grain yield (g/ha)	Single plant yield (g)	No. of pods/plant	100-grain weight (g		
Genotype (G)	19	15.96**	3.08**	129.48**	13.01**		
Env. + (Geno x Env.)	100	42.44**	5.34**	128.51**	3.42**		
Env. (Lin.)	1	3803.81**	410.81**	8642.29**	224.94**		
Gen. x Env. (Lin.)	19	- 5.32++	3.38**	90.44**	1.86*		
Pooled dev.	80	4.23++	0.74++	31.13++	1.02+		
Pooled error	228	1.98	0.22	8.70	0.56		

^{*} and ** significant at 5 and 1 per cent probability level respectively when tested against pooled deviation.

The linear component of G x E interaction was higher in magnitude than non-linear component. The deviation from linear response of variety (pooled deviation) was found significant for all characters. Mehra and Ramanujam (1979) and Govil (1981) reported that a large portion of G x E interaction was accounted for by the linear regression although non-linear component of G x E interaction was also significant in gram for four characters.

Phenotype may be defined as a linear function of genotype (G), Environment (E) and G x E interaction effect. Relative importance of main and interaction effects may vary from genotype to genotype and with the environments. Thus, the study of G x E interactions serves as a guide and help in identifying suitable genotypes for various environmental niches. Finlay and Wilkinson (1963) considered linear regression as a measure of stability, whereas, Eberhart and Russell (1966) emphasised that both linear (bi) and non-linear (Sd-2) components of interaction be considered while judging the phenotypic stability of a genotype. From subsequent studies on this aspect, it is suggested (Paroda and Hayes, 1971) that the linear regression (bi) could simply be regarded as measure of response of a particular genotype whereas, deviation from regression (Sd-2) should be considered as measure of stability. Individual genotypes have been discussed in the light of above information for different characters.

Pods/plant

An examination of stability parameters (bi) and Sd² for the number of pods per plant showed that fifteen genotypes had unit regression value, three had bi value greater than one and two genotypes had bi value below one. Thus, these

genotypes may be categorised as average, abov average and below average sensitive respectively SGM 84-117 and SGM 84-120 having significantly high regression value indicated very high respons had high mean performance suggesting thereb better performance of these genotypes in bette environments (normal sown). However, thesgenotypes were found highly unstable. In contract to these, variety SG 2 showed highest mean, abov average response and was found stable. SGN 84-113 had above average performance, average response and was found stable. However, Jain e al., (1984) reported that none of the genotypes had high pod number combined with unit regression and non-significant deviation from regression. Ir study, varieties C 235, SGM 84-154, IH 83-19, IH 83-18, IH 83-10, and IH 83-9 had below average performance, average response and were found to be stable.

100 grain weight

Hundred grain weight varied from 11.50 (IH 83-6) to 17.60 (SGM 84-104). The genotypes SGM 84-104 and IH 83-10 having relatively bolder seed size had above average response but were found highly unstable. Singh and Choudhary (1980) in soybean reported that varieties with boldest seed measured by 100-seed weight, were most suited for growing in favourable environment. Tomer et al. (1973) while studying phenotypic stability in bengal gram concluded that large seeded cultivars were phenotypically more unstable than small seeded ones.

Yield / plant

Thirteen genotypes had average (bi = 1), three had above average (bi> 1) and four had below average response (bi<1). SGM 84-117 and SGM

⁺ and ++ significant at 5 and 1 per cent probability level respectively when tested against pooled error.

Table 2. Estimates of stability parameters for yield and its components

Genotypes		No. of pods/plant			100-grain weight			
	x	bi	Sd ⁻²	$\overline{\mathbf{x}}$	bi	Sd ⁻²		
IH 83-4	28.6	0.492	1.903*	11.9	0.366*	0.097		
TH 83-6	34.0	0.879	12.768*	11.5	0.759	0.086		
IH 83-9	27.2	1.087	2.742	14.9	0.789	.390**		
IH 83-10	19.1	0.509	8.581	16.6	1.742*	.690**		
IH 83-14	30.3	1.530	26.797**	13.7	1.199	0.043		
IH 83-16	28.2	0.379*	65.623**	12.9	0.487	0.329		
IH 83-18	24:2	0.675	11.697	13.2	0.924	0.259		
1H 83-19	28.2	0.566	4.626	13.7	0.732	0.138		
TH 83-23	24.7	0.326*	69.426**	13.9	1.051	0.485		
IH 83-62	31.5	1.054	16.950*	12.8	0.561	0.011		
IH 83-103	31.8	1.172	24.484**	14.2	0.812	0.205		
SGM 84-104	32.5	0.985	25.032**	17.6	1.905**	.622**		
SGM 84-112	34.7	1.119	. 14.171*	14.1	1.676*	.146*		
SGM 84-113	34.4	1.550	12.077	14.1	0.905	0.073		
SGM 84-117	34.6	1.726*	78.384**	15.1	1.282	0.179		
SGM 84-120	31.0	1.629*	23.920**	14.6	1.049	1.897*		
SGM 84-154	29.3	1.128	2.876	13.9	1.149	0.431		
BR 77	27.2	0.485	26.603**	12.6	0.781	0.466		
C 235	28.7	1.005	6.607	12.8	0.831	0.514		
SG 2	38.8	1.704*	-4.065	12.6	0.897	0.263		
Mean	30.0			13.80		-		

^{*} and ** significant at 5 and 1 per cent probability level respectively.

Genotypes		Single plant yield		Grain yield			
	x	- bi	Sd ⁻²	x	bi -	Sd ⁻²	
IH 83-4	4.9	0.592*	0.465*	18.2	1.220	-0.534	
1H 83-6	5.6	0.592*	0.253	19.5	0.860	-0.287	
IH 83-9	5.4	1.312	0.332*	17.1	1,076	-0.295	
IH 83-1C	4.1	0.623	0.045	14.4	0.864	3.243	
IH 83-14	5.0	1.341	0.338*	15.9	1.262	1.453	
IH 83-16	5.1	0.748	0.913**	18.2	1.260	3.882*	
IH 83-18	4.2	0.727	-0.022	15.1	1,143	-0.552	
IH 83-19	5.3	0.792	0.107	18.1	0,869	1.760	
#H 83-23	4,6).323**	1.611**	17.1	1.035	1.979	
IH 83-62	5.4	0.831	1.221**	. 19.0	1.236	8.782*	
IH 83-103	5.9	1.450	0.169	17.3	0.801	0.733	
SGM 84-104	6.7	1.357	-0.107	20.5	0.836	0.507	
SGM 84-112	6.5	1.256	0.472*	20.9	1.027	4,933*	
SGM 84-113	5.9	1.308	0.295	17.8	0.763	-1.552	
SGM 84-117	6.5	.761**	1.464**	18.8	0.791	0.605	
SGM 84-120	5.9	.660**	1.872**	17.3	0.992	3.422*	
SGM 84-154	5.1	0.870	0.109	17.5	1.042	0.077	
BR 77	5.0	0.560*	1.104*	17.8	0.991	3.606	
C 235	4.92	0.810	-0.022	16.2 -	0.829	-0.636	
SG 2	5.8	1.080	-0.148	19.3	1.103	-2.437	
Mean	5.4	-	1	17.8			

^{*} and ** significant at 5 and 1 per cent probability level respectively.

And the part of the second	State Transfer	N. C.				
			Correlations between			
Characters	₹ vgbi	₹ Vs S ⁻² d	bi Vs S ⁻² d		of yield Vs bi of all components.	S ⁻² d of yield Vs S ⁻² d of yield components
No. of pods/plant	0.751**	-0.049	-0.128	-	-0.182	0.233
100-grain weight	0.802**	0.738**	0.659**		-0.362	0.200
Single plant yield	0.727**	0.094	0.097		0.198	0.526**
Grain yield	0.070	0.131	0.316			$\alpha + \hat{\gamma}_1$

Table 3. Correlations between stability parameters and between stability parameters of yield and its components

84-120 had high mean, above average response but highly unstable for yield per plant. The highest yielding genotype SGM 84- 104 was found to have average response and highly stable. Another genotype IH 83-103 was high yielder having very high response and stable, thus this genotype is likely to perform better in all environments. Variety C 235 and SG 2 showed average response and were highly stable for yield per plant. Mehra and Ramanujam (1979) also reported that variety C 235 was stable for yield per plant with average response.

Grain yield/ha

Grain yield varied from 14.40 (IH 83-10) to 20.90 q/ha (SGM 84- 112). The genotype SGM 84-112, having highest yield was found unstable, however, the genotypes SGM 84-104, IH 83-6, and SG 2 which ranked second, third and fourth in respect of yield were found highly stable. Similar results were reported by Mehra et al. (1980) in gram. Considering the Sd-2 it was found that five genotypes were unstable for grain yield as they had significantly high Sd⁻² values. Promising lines for grain yield which were found stable were SGM 84-117, IH 83-4, IH 83-19, and SGM 84-113. Variety C 235 showed below average yield but highly stable.

. There was lack of general association between mean (X) and stability (Sd-2) and between responsiveness (bi) and stability (Sd-2) for all characters except 100-grain weight (Table 3). This indicated that the separate genetic systems were perhaps involved in the control of those parameters. In contrast to this, there was positive and significant correlations between mean performance of the genotypes (X) and responsiveness (bi) for number

of pods/plant, 100-grain weight and single plant yield. It revealed that genotypes with high mean values for these attributes were, in general, better responsive. There was lack of association between responsiveness (bi) of yield and its components and between stability of (Sd-2) yield and components. This indicated that yield components alone cannot be considered as index for the stability of yield. However, signle plant yield can be taken as index for the stability of yield as such which wa indicated by positive and significant correlation between stability of yield per plant and yield pe plot.

REFERENCES

EBERHART, S.A. and RUSSELL, W.A. (1966).Stability parameters for comparing varieties. Crop Sci., 6: 36-40.

FINLAY, K.W. and WILKISON, G.N. (1963). The analysis of adaptation in plant breeding programme. Austr. J. Agr. Res., 14: 752-754.

GOVIL, J.N. (1981). Analysis of adaptive response in chickpea collection. Legume Res., 4: 23-26

JAIN, K.C., PANDYA, B.P. and PANDE, K. (1984). Stability of yield components of chickpea genotypes. Indian J. Genet., 44: 154-163.

MEHRA, R.B. and RAMANUJAM, S. (1979). Adaptation in segregating populations of bengal gram. Indian J. Genet., 39: 492-500.

MEHRA, R.B., BAHL, P.N. and PAHUJA, A.M. (1980). Phenotypic stability and adaptability in newly developed varieties of chickpea in northern India. Indian J. Agric. Sci., 50: 218 - 222.

PARODA, R.S. and HAYES, J.D. (1971). An investigation of genotype environment interactions for rate of ear emergence in spring barley. Heredity 26: 157-178.

SINGH, O. and GHOUDHARY, B.D. (1980). Stable genotypes for boldness in soybean. Madras Agric. J., 67: 669-670.

TOMER, G.S., SINGH, L., SHARMA, D. and DEODHAR, A.D. (1973). Phenotypic stability of yield and some seed characteristics in bengal gram (Cicer arietinum L.) varieties. JNKVV Res., 7: 35-39.

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^{**} Significant at 1% level.