percentage as compared to control (27.83) and these mutants also showed higher starch percentage. An improved cooking quality was observed in many of the mutants except in CPM 49 and 61, as against the control. Microscopic observation on starch grain size presented in Table 2., also showed variability and the maximum starch grain size observed was 15.0 u in CPM. 46 and against 10.8 u in control. Data presented on reducing sugar showed both increasing and

decreasing trends in mutuant as compared to their control.

ACKNOWLEDGEMENT

The authors are grateful to the Director, CTCRI, Trivandrum for providing the necessary facilities for undertaking the work. The authors are also thankful to Dr. S.N. Murthy, Scientist S-2, CTCRI, for his kind help in Biochemical analysis.

REFERENCES

Ramachandran, K. 1967, Cytology of the genus coleus. Cytologia. 32(3): 474-480.

Upadhya, M.P. and A.N. Purohit, 1973.
Mutation induction and screening procedure for physiological efficacy in potato. In Induced mutations in vegetaively propagated plants. IAEA, Vienna, pp. 61-66.

Madras Agric. J. 79, (3): 138 - 141 March. 1992

Vasudevan, K. and J.S. Jos, 1988. Gamma ray induced mutants in Colcus. Mutation Breeding Newsletter. 32(1988) p.5.

Vasudevan, K., J.S. Jos and M. Unnikrishnan. 1988 b. Remodelling of phenotype in tuber crops through induced mutations. National Symp. on New Trends in Biotechnology, Jone 3, 4 (1988), Trivandrum p. 35 (Abst.).

https://doi.org/10.29321/MAJ.10.A01744

BIOCONVERSION OF RICE STRAW INTO PROTEIN RICH FEED

A. THANIKACHALAM and M. RANGARAJAN

ABSTRACT

Three cellulolytic fungi were inoculated on alkali hydrolysed rice straw and untreated rice straw. Asperquillus sp. (AS.3) exhibited maximum straw conversion (80.0%) followed by Fusarium sp. (Fs.4) and Trichoderma sp. (T.3) Fungal biomass yield ranged from .82 to 1.06 g/50 ml depending upon the fungi. Crude protein content of rice straw after

Centre of Advanced Studies in Agricultural Microbiology Tamil Nadu Agricultural University, Coimbatore 641 003

fermentation varied from 15.6 to 25.0% irrespective of the type of rice straw and the fungi. Soluble protein of the fermented rice straw was maximum (42.5 mg/100 ml) in Aspergillus sp. (As.3) insulated substrate. The results showed the possibility of obtaining protein enriched rice straw employing efficient cellulosic fungi.

Availability of rice straw as a large cellulosic waste (Vander Wal, 1979) has led to the development of suitable fermentation process to convert waste materials into protein rich food or feed. Production of fungal protein from sugarcane bagasse and groundnut shell with Aspergillus niger (5 strains), Penicillium chrysogenum (S.T.F.38) and Pastalotia sp. was reported (Sitaram et al., (1978). Semisolid fermentation of rice and wheat straw acid hydrolysates with Penicillium funiculosum and Candida utilis resulted in 37-180% increase include protein content within 5-7 days at room temperature. Bioconversion of alkali treated rice straw with Myrothecium varrucaria gave the maximum yield of 32.1% crude protein (Dhillon et al., 1981). We recently reported the possible protein enrichment of rice straw through fermentation with Cellulomonas sugar release from rice straw inoculated with certain fungi (Thanikachalam and Rangarajan, 1986 a & b) and scaling up of single cell protein from rice straw by bacterial fermentation (Thanikachalam and Rangarajan, 1987). The present study deals with the bioconversion of rice straw into protein rich fee by three cellulolytic fungi.

MATERIALS AND MEHTODS

Three cellulolytic fungi viz., Trichoderma sp. (T.3), Aspergillus sp. (Ag.3) and Fusarium sp. (Fs.4) isolated from decomposing rice straw, farm yard manure and compost respectively, were used in the present study. The cultures were maintained on potato dextrose agar (PDA) at 10°C.

- (i) Preparation of Pre-treated Rice straw: Rice straw was dried, milled (powdered) and pre-treated as follows: A quantity of 40 g of powdered rice straw was boiled with 750 ml of alkali mixture (5% NaOH and 1% H₂O₂ in 2:1 ratio v/v) for an hour at 100°C. The powder was washed to free the alkali, and sieved (710).
- (ii) Preparation of Inoculum: Fungi were grown under static conditions on Czapek's medium with 0.5% glucose and 0.5% lactose as described by Thanikachalam and Rangarajan (1986 b). After 8 days of growth, fungal mat was homogenised in waring blender and diluted with sterile distilled water to contain 4 x 10⁶ spores/ml. Each flask was inoculated with 2 ml inoculum per 100 ml of medium.
- (iii) Rice Straw Fermentation: Fungi were cultivated in 50 ml. of the broth medium (Reese and Mandels, 1959) dispensed in 500 ml. Erlenmeyer flask with 1% straw powder as carbon source. The flasks were inoculated as mentioned above and were incubated at static condition for 25 days. Per cent conversion of straw into biomass was estimated as follows: Fungal mycelium along with undigested rice straw powder, was filtered

and were transferred to silica crucible. The residue was dried at 60°C and weighed. Then the fungal mat was digested with 100 ml of 5-10% NaoH by keeping in shaker for 6 days and then neutralised by adding con. H2 SO4 drop by drop. The unutilized straw residue was filtered, dried and weighed as mentioned earlier. Control without inoculation was treated similarly and value deducted. In another set of experiment fungi were grown in 100 ml of same medium dispensed in 500 ml conical flask for 25 days under static condition. The residue consisting of fungal mat and unutilised straw, was separated, dried and weighed.

Analytical Methods: The crude protein content of the residue was estimated by micro-Kjeldahl's method (Humphries, 1956). Soluble protein (Lowry et al. (1951) and Cellulase (C_x) activity were determined in the culture filtrate.

RESULTS AND DISCUSSION

Increased conversion of rice straw and biomass yield were observed when treated straw was grown with any one of the three fungi (Table 1) Aspergillus sp. (Ag.3) exhibited maximum per cent of conversion (80.0) followed by Fusarium sp. and Trichoderma sp. (78.0% and 76.0% respectively. But in untreated straw, Fusarium sp. (Fs.4) followed by Trichoderma sp. (T.3) and Aspergillus sp. (Ag.3) recorded maximum conversion of 74.0%, 62.0% and

TABLE 1. Per cent conversion of rice straw and biomass yield by cellulolytic fungi

Cellulolytic fungus		ent of onversion	Biomass yield (g/50 ml)		
	Treated	untreated	treated	untreated	
Trichoderma sp.(T3)	76.00	62.00	0.82	0.66	
Aspergillus sp.(Ag.3)	80.00	52.00	1.06	0.75	
Fusarium sp. (Fs.4)	78.00	74.00	0.93	0.80	

TABLE 2 Growth and activity of cellulolytic fungi on treated and untreated rice straw

Cellulolytic Fungus	Soluble Protein ¹		Cellulolytic, activity ² C		Residue ³		Crude protein in residue (%) ⁴	
	Т	UT	T	UT	T	UT	T	UT
Trichoderma sp. (T.3)	31.80	39.70	78,00	48.00	1.63	1.73	21.80	17.50
Aspergillus sp. (Ag.3)	42.50	43.90	81.00	78.00	1.52	1.61	18.75	15.60
Fusarium sp. (Fs.4)	24.50	39.50	54.00	45.00	1,67	1.89	25.00	19.40

^{1 =} Soluble protein in culture filtrate mg/100 ml.

^{4 =} Crude protein content of residue

^{2 =} Per cent loss in viscosity at 30 minutes

T = Treated Straw

^{3 =} Undigested straw + mycelial mat g/100 ml

UT = Untreated Straw

52.0% respectively. Residue yield containing both fungal mat and undigested straw was found to be more in *Fusarium* sp. (Fs.4) followed by *Trichoderma* sp. (T.3) and *Asperquillus* sp. (Ag.3) Regarding crude protein content of residue, no significant difference between treated and untreated straw as well as among the three fungi was observed (Table 2).

Similar observation was also made for Aspergillus sp, grown on pretreated paddy straw (Ghose, 1981). Maximum crude protein content of 25.0% in treated straw grown with Fusarium sp. (Fs.4) was in agreement with earlier reports (Christias et al. 1975). All the three fungi gave more C_x activity in treated straw than in untreated straw.

REFERENCES

- CHRISTIAS, C., GOVARAKI, C., GEOR-GOPOULOS, S.G., MACRIS, B., and VOMVOYANNI, V., 1975. Protein content and amino acid composition of certain fungi evaluated for microbial protein production. Appl. Microbiol., 29: 250-254.
- DHILLON, G.S., KALRA, K.L., SINGH, S.K., KAHLON, S.S., and KALRA, M.S., 1981. Bioconversion of the delignified rice straw by cellulolytic fungi. In Recycling Residues of Agriculture and Industry (Ed., M.S., Kalra) Bico printers. Ludhians: 77-85.
- GHOSE, T.M., 1981. Cellulosic conversion.
 In: Advances in food producing systems for Arid and Semi-arid land.
 Part A1 (Ed.J.T. Manessabr and E.J. Briskey). Academic Press, London, 225 266.
- HUMPHRIES, E.C., 1956. Mineral components and ash analysis. In: Modern methods of plant Analysis. Springer Verlag, Berlin, 1: 468-502.
- LOWRY, O.H., ROSEBROUGH, N.J., FARR, A.L., and RANDAL, R.J., 1951. Protein measurement with the folinphenol reagent. J. Biol. Chem. 193: 265-275.

- REESE, E.T. and MANDELS, M., 1959.
 B-1, 3-glucanases in fungi. Can.j.
 Microbiol., 5: 173.
- SITARAM, N., MOHAMMAD KUNHI,
 A.A., GEETHA DEVI, B.R., and
 RAMACHANDRA RAO, T.N., 1978.
 Production of fungal protein from
 cellulosic plant materials. Indian J.
 Microbiol., 18: 90-92.
- THANIKACHALAM, A., and RAN-GARAJAN, M., 1986 A. Protein encichment of rice straw with Cellulomones sp. Agricultural wastes, 15 (3): 215-224.
- THANIKACHALAM, A. and RAN-GARAJAN, M. 1986 b. Sugar release from paddy straw inoculated with mixed culture of three cellulolytic fungi. Indian J. Microbiol., 26: 113-116.
- VANDER WAL, P. 1979. SCP for animal feeding. In Proc. fifth International congress of food Sci. Technol., (Ed. E. Chiba, M. Fujimaki, K. Jwai, H. Mitsuda and Y. Morita). Elsevier Scientific publishing company, New York: 47-57.