

## Studies on Induced Chlorophyll Mutations in *Cajanus cajan* (L.) Mill. Sp.\*

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In the study of induced chlorophyll mutations with two varieties of redgram (*Cajanus cajan*), the frequencies increased with increasing doses of mutagens up to middle doses and thereafter decreased. Multiple chlorophyll mutants comprising two types were more frequent than single and three types. The pooled segregation ratios did not show any dose dependency. None among the segregating progenies showed the theoretical expected proportions of 25 per cent of recessive mutants. The spectrum of chlorophyll mutants consisted of *albina*, *xantha chlorina* and *viridis*. *Chlorina* and *viridis* were the predominant types in gamma rays DES treatments respectively.

The use of induced mutations has now come of age as one of the methods of plant breeding. In *Cajanus cajan* attempts on artificial induction of mutations have been made only recently. An attempt has been made in this paper to study the induced chlorophyll mutations in this important grain legume.

### MATERIAL AND METHODS

Two redgram varieties SA.1 and Co.2 formed the material for the study. Gamma irradiation was carried out using a 60 CO Gamma Cell with the doses of 1, 5, 10, 15, 20, 25, 30 and 40 krad. Treatments with chemical mutagen (Diethyl sulphate) were performed by keeping the pre-water soaked seeds immersed for 6 hours in the concentrations of 3, 6, 9, 12 and 15 mM of DES with intermittent shaking. The

DES solutions were changed for every half an hour with a freshly prepared one. Treatments were at room temperature,  $23 \pm 1^\circ\text{C}$ .

After mutagenic treatments,  $M_1$  generation was raised. The  $M_2$  generation was raised as individual  $M_1$  plant and  $M_1$  branch progeny bases. The  $M_2$  seedlings were observed from the day of germination up to 15th day to spot out chlorophyll mutants which were classified according to the system proposed by Gustafsson (1940) and Blixt (1961). The frequency of mutation per 100  $M_1$  plant and 100  $M_1$  branch were calculated from the data. The total number of  $M_2$  mutants and normal seedlings were counted from all the families (both segregating and non-segregating) to calculate the frequency of mutation per 100  $M_2$  seedlings. The different types of seedling muta-

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tions for chlorophyll deficiency were scored separately for calculation of the spectrum of mutants.

## RESULTS AND DISCUSSION

a) *Mutation frequency*: The chlorophyll mutation frequencies estimated as number of mutations per 100 M<sub>1</sub> plants, 100 M<sub>1</sub> branches and 100 M<sub>1</sub> seedlings are presented in Table 1. The trend in mutation frequency on M<sub>1</sub> plant and M<sub>1</sub> branch bases was more or less similar and they did not show any relationship with increase in doses of both mutagens in both varieties. The frequencies of mutations are usually estimated on M<sub>1</sub> plant M<sub>1</sub> spike and M<sub>2</sub> plant bases. Among the three, the third method proposed by Gaul (1960) is preferred as it is proportional to the initial mutation rate and is rather independent of variation in progeny and size of mutated sector. Mohan Rao (1972) also expressed the same view from his study with barley. In the present investigation, in SA.1, the frequency of mutation on M<sub>2</sub> seedling basis increased upto 15 krad of gamma rays and upto 9 mM of DES beyond which there was a decrease. In Co.2, the frequency on M<sub>2</sub> seedling basis increased upto 10 krad of gamma rays and 6 mM of DES after which, there was a decrease. Similar results were reported by Ramasamy (1973) in blackgram and Vindhivarman *et al.* (1980) in *Vigna marina*. The reduction in frequency at higher doses has been attributed to the rigour of both diploptic and haploptic selections in the biological material (Swaminathan, 1961) and elimination of mutations.

b) *Frequency of single and multiple mutations*: The data on relative percentage of M<sub>1</sub> plant progenies segregating for single and multiple mutant types are presented in Table 1. In both the varieties, after gamma rays and DES treatments, multiple chlorophyll mutations comprising two types were more frequent than single and three types. This agreed with the results reported by Muthiah (1975) in sesame. In general, the occurrence of multiple mutations did not bear any consistent relationship with doses of mutagens in both the varieties.

c) *Pooled segregation ratio*: The pooled segregation ratio of chlorophyll mutants in the M<sub>2</sub> generation estimated on M<sub>1</sub> branch basis was higher than those of M<sub>1</sub> plant basis following gamma rays and DES treatments in both the varieties (Table 2). The ratios did not show any dose dependency on both M<sub>1</sub> plant and M<sub>1</sub> branch progeny bases. Such an inconsistent relationship with doses was reported after gamma rays and EMS treatments by Sundaram (1974) in redgram. All the segregating M<sub>1</sub> plant and M<sub>1</sub> branch progenies showed less than 25 per cent of recessive mutants in treatments with both mutagens in both the varieties. The reason for not realising the expected segregation ratio of 25 per cent might probably be due to the induced genic changes with a low transmission frequency closely associated with certain minute chromosomal aberration (deletions) which alter the gene transmission as reported by Moh and Alan (1968) with studies on *Phaseolus vulgaris*. The small mutated

sector involving more number of initial cells and lethality of mutated cells may be the other cause for the realisation of a lower segregation ratio.

d) *Mutation spectrum*: The spectra of mutants observed in the segregating progenies were estimated and their relative percentages are given in Table 3. In both the varieties, the spectra of mutants comprised of only four types namely, *albina*, *xantha*, *chlorina* and *viridis*. Alikhan and Veerasamy (1974) also reported the occurrence of four types only in the same crop. In both the varieties, *chlorina* was the most frequent type following treatments with gamma irradiation, whereas *viridis* followed by *xantha* occurred in a high proportion after DES treatments. Alikhan and Veerasamy (1974) reported the frequent occurrence of *chlorina* types in gamma rays treated population of redgram. Gustafsson (1963) and Nilan (1972) reported that alkylating agents induced high proportion of *viridis* and low proportion of *albina*. Westergaard (1960) found that the alkylating chemicals induced the less drastic mutations such as *viridis* instead of extreme mutations such as *albina* in higher proportions than those induced by radiations possibly because of their apparently less drastic effect on chromosomes. A maximum of three types of mutants were recorded in the individual doses of gamma rays or DES in both the varieties. The occurrence of mutant types, however, was not proportional with the doses of mutagens. Similar inconsistent result with doses of mutagens was reported by Vindhiyavarman *et al.* (1980) in *Vigna marina*.

## REFERENCES

- ALIKHAN, W. M. and R. VEERASAMY 1974. Mutations induced in redgram (*Cajanus cajan* (L.) Mill sp.). *Radiat Bot.* 14: 219-22.
- BLIXT, S. 1961. Quantitative studies of induced mutations in peas, V. Chlorophyll mutations. *Agri. Hort. Gen* 18: 219-27.
- GAUL, H. 1960. Critical analysis of the methods for determining the mutation frequency after seed treatment with mutagens. *Genet. Agr.* 12: 297-18.
- GUSTAFSSON, A. 1940. The mutation system of the chlorophyll apparatus. *Lunds. Univ. Arsskr.* 36: 1-40
- GUSTAFSSON, A. 1963. Productive mutation induced in barley by ionizing radiations and chemical mutagens. *Hereditas.* 50 : 211-63.
- MOH, C. C. and J. J. ALAN. 1968. Comparative study of the mutation frequency and genetic behaviour mutants induced by acute and chronic gamma irradiation in common beans (*Phaseolus vulgaris* L.). *Aplicaciones de la energia nuclear al aumento de la Prod. agricola.* Union panamericana, washington
- MOHAN RAO, P. K. 1972. The relative merits of the three methods of measuring mutation frequency in barley. *Radiat. Bot.* 12:323-29. 85-93.
- MUTHIAP, K. N. 1975. *Induced mutagenesis in diploid and autotetraploidsesame (Sesamum indicum L.)*. M.Sc. (Ag.) Dissertation (Unpubl.). Tamil Nadu Agri. Univ., Coimbatore.
- NILAN, R. A. 1972. Mutagenic specificity in flowering plants. Facts and Prospects. *Induced mutations and Plant improvement*. STI/PUB/ 297; IAEA, Vrenna, 141-151.
- RAMASAMY, N. M. 1972. Investigations on induced mutagenesis in blackgram (*Phaseolus mungo* L.). Ph.D. Thesis? (Unpubl.), Tamil Nadu Agri. Univ., Coimbatore.
- SUNDARAM K., MOHANA. 1974. Induced mutagenesis in redgram *Cajanus cajan* (L.) Mill Sp.) M.Sc. (Ag.) Dissertation (Unpubl.). Tamil Nadu Agri. Univ., Coimbatore.

SWAMINATHAN, M. S. 1961. Effect of diplo-  
 tic selection on the frequency and spectrum  
 of mutations induced in polyploids following  
 seed irradiation. *Symp. on the effects of  
 ionizing radiation on seeds*. IAEA, Vienna,  
 279-88.

VINDHIYAVARMAN, P., CHANDRASEKARAN, P.  
 and KRISHNASAMY, S. 1980. Induced  
 chlorophyll mutations in *Vigna mungo* (Burm.)  
*Morr. Madras agric. J.* 67: 425-28.

WESTERGAARD, M. 1960. A discussion of  
 mutagen specificity. *Chemische mutagenese*.  
 Erwin—Bauer Gedächtnis Voleumgen I: Acad.  
 emic—Verlag, Berlin, pp. 116-21.

Table 1 Mutation frequency and relative % of multiple mutations

Mutagen and dose	Chlorophyll mutation frequency						Relative % of M <sub>1</sub> plant progenies segregating for mutations of					
	Per 100 M <sub>1</sub> pinats		Per 100 M <sub>1</sub> branches		Per 100 M <sub>1</sub> seedlings		One type		Two type		Three type	
	SA. 1	CO. 2	SA. 1	CO. 2	SA. 1	CO. 2	SA. 1	CO. 2	SA. 1	CO. 2	SA. 1	CO. 2
<b>I Gamma rays (krad)</b>												
1	28.9	24.7	9.4	10.2	1.8	1.5	11.5	16.7	88.5	83.3	—	—
5	28.4	28.9	19.8	19.1	2.4	2.8	14.3	—	85.7	88.5	—	—
10	17.1	17.8	16.1	16.4	3.0	4.1	16.7	—	66.6	100.0	16.7	—
15	18.0	24.8	17.6	23.6	3.7	3.8	—	23.5	100.0	76.5	—	—
20	16.7	15.0	15.7	13.8	2.2	2.3	—	16.7	100.0	68.6	—	—
25	14.8	14.3	11.1	11.5	1.2	1.0	—	20.0	75.0	80.0	25.0	—
30	18.2	22.2	12.5	17.3	1.1	1.4	—	—	100.0	100.0	—	—
40	25.0	20.0	10.5	16.7	0.7	0.9	—	—	100.0	100.0	—	—
<b>II DES (mM)</b>												
3	25.0	26.4	25.5	12.5	1.2	1.6	18.2	17.4	81.8	82.6	—	—
6	24.0	29.2	16.5	24.7	2.2	3.5	—	14.3	100.0	76.2	—	9.5
9	45.2	22.2	26.9	20.6	2.9	2.6	10.5	—	73.7	100.0	15.8	—
12	36.8	28.1	21.6	20.0	1.6	1.1	14.3	11.1	85.7	88.9	—	—
15	—	25.0	—	20.7	—	0.8	—	33.3	—	63.7	—	—

Table 2. Segregation percentages of chlorophyll mutants in  $M_3$  generation

Mutagen and dose	Total No. of $M_1$ plants segregating		Pooled segregation ratio (%)		Total No. of $M_1$ branches segregating		Pooled segregation ratio (%)	
	SA. 1	CO. 2	SA. 1	CO. 2	SA. 1	CO. 2	SA. 1	CO. 2
<i>I. Gamma rays (krad)</i>								
1	26	24	13.7	11.7	28	26	15.1	13.1
5	28	26	11.8	12.5	34	50	13.4	14.3
10	12	13	12.1	10.6	30	30	14.3	13.5
15	11	17	10.2	11.9	29	42	14.9	13.5
20	7	6	11.8	12.5	18	15	13.1	14.8
25	4	5	10.0	12.5	7	10	13.9	14.5
30	2	4	10.0	10.0	3	9	14.2	13.6
40	2	2	12.5	12.5	2	4	15.0	15.0
<i>II. DES (mM)</i>								
3	22	23	10.2	11.0	53	29	13.0	13.4
6	12	21	10.4	11.8	20	44	14.5	14.4
9	19	12	13.8	11.3	28	29	16.0	13.7
12	14	9	9.6	12.5	21	16	12.7	13.8
15	---	3	---	12.5	---	6	---	14.2

Table 3. Spectrum of chlorophyll mutants in M<sub>2</sub> generation

Mutagen and dose	Total No. of mutants in M <sub>2</sub>		Spectrum (Relative percentage) of chlorophyll mutants							
	SA. 1	CO. 2	Albina		Xantha		Chlorina		Viridis	
	SA. 1	CO. 2	SA. 1	CO. 2	SA. 1	CO. 2	SA. 1	CO. 2	SA. 1	CO. 2
<b>I Gamma rays (krad)</b>										
1	217	159	2.3	—	40.1	—	57.6	58.5	—	41.5
5	232	295	—	2.0	32.8	33.2	67.2	64.8	—	—
10	264	384	6.1	—	—	—	58.7	58.6	35.2	41.4
15	304	321	—	—	—	24.6	50.3	21.5	49.7	53.9
20	110	108	—	5.7	—	31.3	22.7	63.2	77.3	—
25	41	38	7.3	—	—	65.8	26.8	—	65.9	34.2
30	19	25	—	—	26.3	—	73.7	40.0	—	60.0
40	4	7	—	14.3	25.0	—	—	85.7	75.0	—
<b>II DES (mM)</b>										
3	110	157	—	—	59.1	56.1	—	—	40.9	43.9
6	154	247	—	1.6	—	36.8	64.3	—	35.7	61.5
9	151	145	3.3	—	29.8	—	—	64.1	66.9	35.9
12	51	33	—	—	39.2	27.3	—	—	60.8	72.7
15	—	8	—	—	—	50.0	—	50.0	—	—