A Fruit Rot of Chillies (Capsicum annuum L.) caused. by Alternaria solani (Ell. & Mart) Jones & Grout

by

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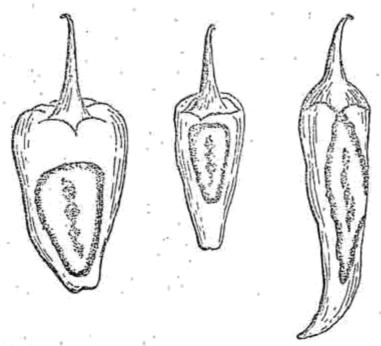
Introduction: Fruit rot of chillies is a disease of economic importance prevalent in many parts of the world. The causal organism differs in different places and more than one fungus has been found to be responsible for this disease. Glomerella cingulata (Stonem.) Spauld. & v. Schrenk (Colletotrichum piperatum Ell. & Ev.) has been recorded from India (Dastur 1920), America (Halsted 1890) and elsewhere causing anthracuose (fruit rot) of Capsicum. Colletotrichum capsici Syd., was first observed in South India, infecting fruits and young twigs of chillies (Mc Rae 1914). Dastur (1921) has described this disease in detail from North India. More than one species of Alternaria have been reported to be responsible for the fruit rot of chillies. A. solani (Ell. & Mart.) Jones & Grout has been recorded from Georgia by Higgins (1925). Wallace (1929) reported a fruit rot of chillies caused by A. tenuis Noes., in Tanganiyka. In Spain, Unamuno (1934) found A. tenuis. f. genuina Una., to be associated with the rotting of chilli fruits. Bremer (1945) from Turkey stated that A. longipes (Ell. & Eve.) Tisd. & Wadk, was the causal agent for the rotting of chilli fruits in that country. A. capsici-annui Savul. and Sandu has been known to be responsible for this disease in Rumania (Sayulescu & Hulea 1948). Dutt (1938) reported a fruit rot of chillies from Delhi, caused by Alternaria but the species was not determined.

In recent years, specimens of chilli fruits affected by Alternaria have been collected from Mathurai and Guntur districts. This disease occurred along with the fruit rot caused by Colletotrichum capsici Syd., and often passed off for infection by the latter fungus. The present paper is concerned with the isolation of the fungus and study of its pathogenicity, cultural characters and host range.

Material and Methods: The pathogen was brought into culture by the single spore isolation method and its growth on a number of agar media was studied. Inoculation experiments were conducted on fruits kept in moist chambers after surface sterilisation or on those growing on the plants themselves. Leaf and twig inoculations were carried out on young plants grown specially for the purpose. For the study of the rate of growth of the fungus, equal quantities of the inoculum were used and the media were adjusted to the same reaction. The colour nomenclature described in Ridgeway's "Colour Standards" has been adopted for describing the colour of the growths on the various media.

Symptoms of the disease: Fruits which were nearing maturity were ordinarily infected. Brown lesions surrounded by a yellow

halo developed on one side of the fruit. These enlarged and resulted in the formation of irregular sunkon patches with a dark brown margin and light grey centre. The latter was overgrown by olive brown fungal growth. The fruit lost its red colour and usually dropped down. Sometimes the entire fruit was involved in the rot.



Chilli fruits showing the symptoms of the disease

The incidence of fruit rot was very high if the humidity increased or it rained during the maturing stages of the fruits, resulting in a heavy loss of fruits. This disease can be distinguished from the fruit rots caused by Colletotrichum capsici Syd. or Glomerella cingulata by the absence of the characteristic acervuli on the diseased spots.

The fungus and its cultural characteristics: To start with, the mycelium is internal and after the collapse of the tissues, brown mycelial growths develop on the surface of the lesions. Numerous light brown, septate, unbranched conidiophores are found in clusters of 3 to 6 from the external mycelium, measuring $105 \times 4\mu$ (90-144 x 3-65). Obelavate, muriform conidia, olive brown in colour are formed, often in chains on the conidiophores and measure $65 \times 18\mu$ (49 to 112×12 to 25) on the average. The number of cells in the spores varies; 8 to 12 transverse septa and 3 to 6 longitudinal septa are found in each spore. The beak is usually small.

Profuse aerial growth usually develops on agar media. In the initial stages, the hyphae are hyaline forming a whitish aerial mycelium. Later, the growth becomes dark grey and forms an olivaceous black mat on the surface of the agar. The dark

discolouration extends into the medium also. Profuse sporulation was evident on most of the media. The cultural characters of this fungus and the average rate of daily growth on various media adjusted to pH 5.7 are given below:

TABLE I Cultural characters on various media

Name of medium		Average rate of daily growth in m.m.	Cultural characters after 7 days	
1.	Ont agar	G:6,	Good growth, aerial mycelium dark grey, olivaceous black on agar surface, roverse black, spores plenty in 48 hours.	
2.	Fronch bean agar	7.5	Fluffy white aerial mycelium, good and uniform growth; the older mycelium tends to be mineral grey, reverse purple black, spores nume- rous within 48 hours.	
3.	Maize agar	53737	Aerial mycolium light mineral grey over a dark dull yellow substratum, roverso sooty black, growth was good and uniform. Spores abun- dant within 48 hours.	
4.	Carrot agar 2 5	7:0	Loose aerial mycelium, pearl to storm grey, adpressed over a dusky olive-green growth on the surface of the agar medium. Zonations noticed on the reverse; reverse black, spores plenty in 48 hours.	
5.	Czapek Dox's agar	7.0	Aerial mycelium white to ash-grey in the centre, on olivaceous black background reverse french grey to light violet grey; no spores in 48 hours.	

French bean agar appears to be the most satisfactory medium for the growth of this fungus.

Pathogenecity: Repeated inoculations were carried out on surface-sterilised fruits of chillies, brinjal and tomato kept in sterilised moist chambers and those growing on plants. Leaves and twigs of Capsicum annuum, Solanum melongena, Lycopersicum esculentum, Datura stramonium, Nicotiana tabacum, Gossypium hirsutum and Cyamopsis tetragonaloba were inoculated. The following table shows the results of inoculation.

TABLE II.
Results of Inoculation with the Fungus.

	Name of plant	Part inoculated	Method of inoculation	Number inoculated	Number infected
l	Capsicum annuum	Leaves and twigs	Wounded	12 plants	Nil
		Leaves and twigs	Unwounded	12 plants	Nil
		Fruits	Wounded	24 fruits	15 fruits rotted within 7 days after inoculation
		Fruits	Unwounded	24 fruits	14 fruits rotted within 7 days after inoculation
2	Solanum melongena	Leaves and twigs	Wounded	6 plants	No infection
		Leaves and twigs	Unwounded	6 plants	No infection
		Fruits	Unwounded	12 fruits	10 fruits rotted within 7 days after inoculation
},	Lycopersi- cum esculen- tum	Leaves and twigs	Wounded	6 plants	No infection
		Leaves and twigs	Unwounded	6 plants	No infection
		Fruits	Wounded	6 fruits	4 fruits rotted within 7 days after inoculation
	.*1	Fruits	Unwounded	6 fruits	3 fruits rotted within 7 days after inoculation
ŧ.	Datura stramonium	Leaves and twigs	Wounded	6 plants	No infection
			Unwounded	6 plants	No infection
5.	Nicotiana Leaves and tabacum twigs		Wounded	6 plants	No infection
			Unwounded	6 plants	No infection
3.	Gossypium hirsutum	Leaves and twigs	Wounded	6 plants	No infection
		2 1 2	Unwounded	6 plants	No infection
7.	Cyamopsis tetragonaloba	Loaves and twigs	Wounded	6 plants	No infection
			Unwounded	6 plants	No infection

Suitable controls were kept in all cases and they remained healthy throughout the experiment.

The results of inoculation show that the fungus is pathogenic on the fruits of chillies, brinjal and tomato. There was no infection on the leaves of any of the plants tried. The symptoms of infection on the fruits were evident on the fourth day after inoculation. The infected fruits were completely rotten in the course of one week. In all cases the fungus was reisolated from the infected fruits.

Identity of the Fungus: Four species of Alternaria have been recorded on Capsicum. A comparative statement of the chief distinguishing characters of the different species of Alternaria recorded on chillies is given below:

TABLE III
Character of the species of Alternaria recorded on Chillies

	Name of species	Conidial measurements in μ	Chief distinguishing _ Authority
1.	A. longipes	30 — 50 × 10 — 13	Infects only wounded chilli Bremer fruits and incapable of 1945 infecting tobacco or chilli leaves
2.	A. solani	145 — 370 × 16 — 18	Infects the leaves of almost Higgins all solanaceous plants 1925
3.	A. tenuis	30 — 36 × 14 — 15	Wallace 1929
4.	A. tenuis f. genuina	27.5 — 62.5 × 12 — 17.5	Found on leaves, peduncles Unamuno floral buds and unripe 1934 - fruits of chillies
5.	A. capsici- annui	33 — 82 × 7 — 21	Capable of infecting tobacco Savulescu and chilli leaves also and Hulea 1948
6.	Alternaria sp.	14 — 50 × 7 — 19	Capable of infecting all por- tions of chillie plant 1938
7,	The present isolate under study	49 — 112 × 12 — 25	Capable of infecting only the fruits of chillies, tomatoes and brinjal

From the above characters of the various species it can be inferred that the isolate under study does not completely agree with any of the others. But, it exhibits affinities to A. longipes in being incapable of infecting tobacco and chilli leaves but infecting the fruits. But the spores are much bigger than those of A. longipes. In this character, it comes near A. solani, but differs from the latter in its failure to infect the leaves of any of the solanaceous

plants tried. It is considered that it is more likely that the present isolate is a strain of A. solani which infects only fruits and not the leaves of chillies, brinjal and tomato.

Control: Diseases caused by Alternaria are amenable to control by Bordeaux spray. Since it is customary to spray chillies for the control of fruit rot caused by Colletotrichum capsici the same treatment will be useful for the control of the disease caused by this fungus.

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BIBLIOGRAPHY

- 1. BREMER, 1945. On pod spot in Pepper. Phytopathology, 35: 283 287.
- DASTUR, J. F., 1920. Glomerella cingulata on chillies and Carica papaya Appl. Biol. 6: 245 - 268.
- 3. 1921. Dieback of chillies in Bihar. Mem. Dept. Agric. India, 11:129-144.
- DUTT, K. M., 1938. Alternaria species on chilli in India. Curr. Sci. 6:97-98.
- HALSTED, B. D., 1890. Rept. Bot. Dept. N. J. Agric. Exp. Sta. Rept. 11: 358-360.
- 6. HIGGINS, B. B., 1925. Blossom end rot of pepper (Capsicum annuum)

 Phytopathology, 15: 223 229.
- MCRAE, W., 1914. Administration Rep. Govt. Mycologist for 1913 '14, Madras Presidency. Rep. on the operations of Dept. Agric. Madras. 1913-14. 50.
- 8. RIDGEWAY, R., 1912. Colour standards & colour nomenclature.
- 9. SAVULESCU, T. & HULEA ANA, 1947. Review of applied Mycology, 27: 458. 1948.
- 10. UNAMUNO, L. M., 1934. Review applied Mycology, 13:84.
- 11 WALLACE, G. B., 1929. Diseases of plants. Rep. Dept. Agric. Tanganyika for the year 1928. 40 42.