

Influence of Nitrification Inhibitor on Green House Gas (GHG) Emission under Intensive Paddy System

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The field experiments were carried out to assess the influence of Dicyandiamide (DCD) along with nine different nitrogen management practices on emission of methane (CH_{2) and} nitrous oxide (N₂O) under intensive paddy system of Cauvery Delta Zone. The CH₂ and N₂O efflux clearly showed daily and seasonal fluctuations. Among the treatments, Leaf Colour Chart (LCC) based N (30 kg N ha⁻¹ keeping the LCC value 4 as standard + DCD @ 10 % of applied N) recorded the lowest average emission of 0.50 mg.m².day⁻¹ and 2.93 mg.m².day⁻¹ of N₂O and CH₃, respectively. Irrespective of all treatments, among the critical stages of crop growth, flowering stage recorded highest emission of 23 % N₂O_{and} 50 % of CH₃when compared to tillering stage. Among the "N" applied treatments, Leaf Colour Chart (LCC) based N + DCD recorded the lowest seasonal green house gas emission (0.49 kg ha⁻¹ & 0.44 kg ha⁻¹ of N₂O and 2.94 kg ha⁻¹ & 3.20 kg ha⁻¹ of CH₄ during Kharif and Rabi seasons, respectively) along with higher paddy yield (12 to 15%).

Key words: Dicyandiamide, Nitrogen fertilizer, Nitrous oxide, Methane, Paddy

Methane and nitrous oxide are the two major Green House Gases (GHG) emitted from paddy (Oryza sativa L.) agro-ecosystem. At global level, rice cultivation alone contributes 10 % of the total CH, emission while the global warming potential of N₂O is 298 times higher (Rees et al. 2013) than carbon di-oxide, so mitigation of both CH, and N,O is needed to combat global warming. In flooded rice, methanogens bacteria consume soil organic carbon and emit CH. In rice soil, NO is produced by both biological (nitrification and denitrification) and chemical decomposition process. Nitrogen based fertilizer is main source of N₂O production in rice soil. Methane and N₂O production in rice soil is influenced by several factors such as water, soil pH, redox potential, temperature, organic matter of soil, soil microbial diversity, transplanting methods, rice cultivar, crop duration and type & time of fertilizer application (Hussain et al., 2015). Application of "N" fertilizers increases N₂O emissions (Bronson and Mosier, 1992). Emissions of N₂O from N-fertilized croplands vary considerably, ranging between 0.001% and 6.8% of applied "N" (Bouwman, 1990; Eichner, 1990). From the agricultural soils, nitrification and denitrification are the two processes responsible for formation of N₂O. In both these processes, nitrite (NO₂-) is formed as an intermediate compound. During the process of nitrification, $\mathrm{NH}_{_{\!4}}{}^{_{\!+}}$, in aerobic condition, gets oxidized to NO3 via hydroxylamine and nitrite, releasing N₂O as a byproduct, while in denitrification, the NO₃ gets completely reduced to N₂ evolving N₂O as an intermediate product. Therefore, the end product of nitrification works as substrate for denitrification. Hence, controlling the first process will certainly help in regulation of second process to ome extent. Nitrification inhibitors are compounds that reduce the rate at which ammonium is converted to nitrate either by killing or interfering with the metabolism of nitrifying bacteria. Dicyandiamide (DCD) is one of the most widely used bacterio-static nitrification inhibitors in the agriculture (Zacherl and Amberger, 1990) and decomposes in soil to non-toxic products. Effect of DCD on N₂O emissions has been reported by Mosier et al. (1996) in wheat and maize and McTaggart et al. (1997) in ryegrass, grassland and spring barley. The present study was undertaken to observe the effect of DCD on N₂O & CH₄ efflux from irrigated rice soils of Cauvery Delta Zone to assess its suitability for decreasing GHG emission to the atmosphere.

Material and Methods

The present investigation was carried out during the year 2013-2014 to assess the influence of Dicyandiamide (DCD) along with nitrogen management practices on the emission of $\rm N_2O$ & $\rm CH_4$ from agricultural soils. A field experiment was conducted at Tamil Nadu Rice Research Institute, Aduthurai (11° N latitude, 79° 31' E longitude, 19.4 MSL). During normal years, the annual rainfall is 1200 mm of which around 70 % is received during September to October (North East Monsoon). The climate of the experimental site (Cauvery Delta) is sub tropical monsoon type. The experiment with fixed plots has been laid out in a Randomized Block Design (RBD) with three replications. The details of the treatments are listed below.

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Treatments

T,: Absolute control

T₂: Blanket recommendation of Nitrogen {150kg N ha⁻¹ in 4 splits 25 % each at basal, 15,30, and 45 Days After Transplanting (DAT)}

T₃: Leaf Colour Chart (LCC) based N (30 kg N ha⁻¹ keeping the LCC value 4 as standard)

T₄: SSNM (Site Specific Nutrient Management) based N with fixed split approach (35 % N at 15 DAT, 40 % N at 30 DAT, 25 % at 45 DAT with use of LCC at each stage)

 $\rm T_{\scriptscriptstyle 5}$: Early completion of N application (25 % basal 50 % at 20 DAT and 25 % at 40 DAT)

T₆: T₂ + Dicyandiamide (DCD) @ 10 % of applied N

 T_7 : T_3 + DCD @ 10 % of applied N

 $T_{_{\rm R}}$: $T_{_4}$ + DCD @ 10 % of applied N

T_a: T₅+ DCD @ 10 % of applied N

*Common application (T_2 to T_9): Each 50 kg ha⁻¹ of Phosphorus & potassium, Micronutrient mixture @ 25 kg ha⁻¹ and Gypsum @ 500 kg ha⁻¹

A uniform plot size of 25 m² was adopted for all the treatments and replications. Nitrogen was applied as per treatment schedule through urea while phosphorus and micronutrient mixture were applied entirely as basal and potassium in two equal splits (basal and at panicle initiation stage). The DCD was applied at the rate of 10% of applied N. Need based plant protection measures were taken up against pest and diseases.

N,O and CH, measurements

The N₂O and CH₄ efflux from all the plots were measured using static chambers during the critical stages of crop growth (tillering, flowering and maturity). To collect the gas generated in the experimental plot, the acrylic chambers (60 cm height with the capacity of 80 lit) were placed on the iron base (47 cm diameter with the total area of 1562 cm²) which was inserted 10 cm inside the soil one day before gas collection. The channel situated at the upper edge of base was filled with water to make the system air tight. One 3-way stopcock (Eastern Medikit Ltd. India) is fitted at the top of chamber to collect gas samples. The chamber should be thoroughly flushed several times with a 50 ml syringe to homogenize the inside air thoroughly. A battery operated fan was run continuously during the sampling to circulate the air inside the chamber to facilitate the proper mixing of the gas inside.

Gas samples were drawn with 50 ml syringes with the help of hypodermic needle (24 gauge) at 0, 10, 20 and 30 min. intervals and syringes were made airtight with a three way stop cock to arrest the gas diffusion. The soil temperature, chamber temperature and water level inside the iron base was recorded during gas collection which was used to calculate N_2O & CH_4 flux. Air samples were brought immediately to the laboratory for N_2O & CH_4 analysis. The N_2O &

 $\mathrm{CH_4}$ concentration was determined by using a gas chromatograph (Varian, 450 GC, German), equipped with Electron Capture Detector (ECD) and Flame Ionization *Detector* (FID), respectively. Under the appropriate operating conditions (column temperature 35°C, injection temperature 120°C and detection temperature 300°C) $\mathrm{N_2O}$ & $\mathrm{CH_4}$ peaks were detected at a specific retention time. Before sample analysis, gas chromatograph was calibrated with different dilutions of 10 ppm of $\mathrm{N_2O}$ & $\mathrm{CH_4}$ gas procured from M/S Multitech Pvt. Ltd., New Delhi, India.

Results and Discussion

The experimental soil is fine, montmorillonitic, isohyperthermic, Udorthentic Chromusterts under the soil textural class 'clay' coming under Kalathur soil series. It was neutral in pH with a soluble salt concentration of < 0.5 dSm⁻¹ (Table 1). The organic carbon status was medium falling in the range of 0.5 to 1 %. Owing to its heavy clay content, it possessed a high cation exchange capacity (42.3 C mol (p+) kg⁻¹).

Table 1. Physico-chemical characteristics of experimental soil

Particulars	Values
Coarse sand (%)	6.60
Fine sand (%)	26.27
Silt (%)	15.5
Clay (%)	47.0
pH (1:2.5)	7.77
Electrical conductivity (dSm ⁻¹) (1:2.5)	0.32
Organic carbon (%)	0.66
Cation Exchange Capacity (C mol (p+) kg ⁻¹)	42.3
Available N (kg ha ⁻¹)	291
Available P (kg ha ⁻¹)	66.0
Available K (kg ha ⁻¹)	234

Influence of different "N" management practices

The measurement of CH, and N,O efflux was carried out from the soils amended with different nitrogen management practices along with 10 % of DCD. It showed daily and seasonal fluctuations. Among the treatments, Leaf Colour Chart (LCC) based N (30 kg N ha¹ keeping the LCC value 4 as standard + DCD @ 10 % of applied N) (T_7) recorded the lowest average emission of 0.50 mg.m² day¹ and 2.93 mg.m².day¹ of N,O and CH,, respectively (Fig. 1).

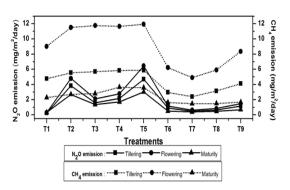


Fig.1. Influence of different "N" management practices on N₂O and CH₄ emission

This might be due to the need based addition of urea under LCC guided N management along with DCD that have probably reduced the net NO₃-N available for denitrification in the soil and it also results in increased competition by plant roots. Irrespective of the days of fertilizer application, the maximum mean emission was recorded as 4.72 mg.m².day¹ of N₂O and 7.13 mg.m².day¹ of CH₄ under early completion of N application (25 % basal 50 % at 20 DAT and 25 % at 40 DAT- T₅) due to higher available NO₃-N.

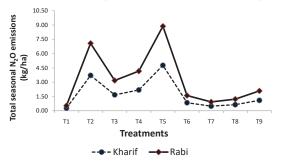


Fig.2. Influence of different "N" management practices on N₂O emission

The peak N₂O efflux observed during this study was associated with application of NH, based fertilizer, i.e. urea. Ammonium based fertilizer application directly stimulates nitrification process, as it serves the substrate for nitrifying bacteria in the oxic conditions. Nitrification inhibitor retards the ammonium mono-oxygenase enzyme which is responsible for converting ammonium present in soil to hydroxyl amine which is further oxidized to nitrite and then to nitrate (Prasad and Power 1995). Initial lower efflux at the start of the experiment was due to the time required for hydrolysis of urea in soil to NH₄. In agricultural fields, increase in N₂O emission typically follows N-fertilization for a short time. After this, emission rates are reduced to fluctuate around a low base line level, independent of the amount of fertilizer applied (Mosier, 1998). A decline in the efflux is attributed to substrate depletion for nitrifying bacteria.

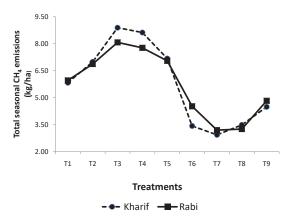


Fig.3. Influence of different "N" management practices on $\mathrm{CH_4}$ emission

Irrespective of all treatments among the critical stages of crop growth, the flowering stage recorded

higher emission of 23 % N_2O_{and} 50 % of CH_a when compared to tillering stage (Fig. 1). This was due to the coincidence of top dressing of nitrogenous fertilizers and active growth stage of crop produced more plant hormones which supported methane emission.

Methane emissions were always lower in the presence than the absence of DCD from our experiments. The application of DCD with basal fertilizer could reduce $\mathrm{CH_4}$ fluxes and hence significantly influenced $\mathrm{CH_4}$ emission. Saturation of soil creates anaerobic conditions conductive for $\mathrm{CH_4}$ emission, as methanogens are strict anaerobes. Therefore, lower inhibitive effect of DCD on $\mathrm{CH_4}$ emission may be found when the DCD applied into soil with tillering and panicle initiation than with basal fertilizer in our experiments.

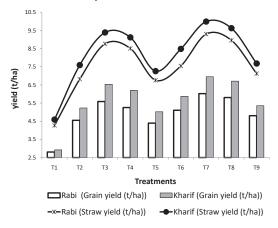


Fig.4. Influence of different "N" management practices on paddy yield

Application of fertilizers along with DCD has been considered to be one of potential mitigation strategies in rice paddy field due to the simultaneous reduction of CH, and N₂O. However, there are conflicting reports regarding the influence of urease and nitrification inhibitors on CH₄ emission (Malla et al., 2005) and the mechanism of urease inhibitors and nitrification inhibitors on CH₄ emission needs to be considered from three different levels. Urease and nitrification inhibitors affected CH₄ emission by the influence on methanogenesis. Wang et al. (1991) indicated that urease inhibitors have little influence on the oxidation of ammonium following the hydrolysis of applied urea in soils, but hydroquinone may decrease CH, production via inhibition of the methanogenic fermentation of acetate.

Among the "N"applied treatments, Leaf Colour Chart (LCC) based N (30 kg N ha¹¹ keeping the LCC value 4 as standard + DCD @ 10 % of applied N) (T_7) recorded the lowest GHG emission of 0.49 kg ha¹¹ & 0.44 kg ha¹¹ of N₂O and 2.94 kg ha¹¹ & 3.20 kg ha¹¹ of CH₄ during Kharif and Rabi seasons, respectively. This might be due to the lower availability of substrate for N₂O emission. The use of DCD reduced N₂O emission significantly by 77-85% and CH₄ emission

by 34-42 % relative to treatments without DCD. Besides increased "N' use efficiency & higher grain yield (6-7%) was observed in all DCD treatments. This is due to, firstly DCD reduces N₂O emissions directly by nitrification (by reducing NH₂ to NO₃) as well as indirectly by de-nitrification (by reducing NO₃ availability in soil). Secondly, methane emission from rice soil can be reduced by enhancing methane oxidation and suppressing methane production and further by reducing the aerenchymal transportation through rice plant. Besides DCD maintained higher soil redox potential, this subsequently reduced the cumulative methane emission. Comparable results were also reported by Kumar and Malyan (2016).

The usage of DCD not only decreasing the GHG emission (30 to 80% reduction of $\rm N_2O$ and $\rm CH_4)$ also increased the crop yield up to 12 to 15% in both kharif and Rabi. Irrespective of the seasons the treatment LCC based N+DCD recorded higher grain and straw yields (grain yield of 6.95 t ha⁻¹ during Kharif , 6.92 t ha⁻¹ during Rabi, straw yield was 9.99 t ha⁻¹ during Kharif and 9.30 t ha⁻¹ during Rabi) followed by SSNM based N+ DCD9G Fig. 4 & 5.).

This is due to the need based application of fertilizers in the treatments considerably increased the crop growth and yield. The DCD applied treatments registered higher grain and straw yields ranged from 5 to 22 % then others. This might be due to; DCD reduced the net NO₃-N available for denitrification in the soil as a result of increased competition by plant roots.

DCD is not a biocide and has no effect on soil microbial biomass (Di and Cameron, 2004). It acts specifically on an enzyme (ammonia monooxygenase) contained in nitrosomonas by blocking the site where ammonia is converted to nitrite. It is also water soluble and biodegradable into carbon dioxide, water and ammonia. Its rate of degradation and its effectiveness decreases with time after application to soils. Increasing the soil temperature, pH, moisture and organic matter content decreases its effectiveness (Irigoyen et al., 2003; Di and Cameron, 2004). DCD is a commonly used nitrification inhibitor; several investigations had also proved that DCD worked as a potential nitrification inhibitor for Indian conditions (Majumdar and Mitra, 2004).

It is naturally broken down in the soil into non-toxic product with no traces of residue left beyond the cropping year. A reduction in the $\rm N_2O$ to the tune of 40% in a dry sandy loam soils has been reported (Skiba *et al.*, 1993), 58–78% when mixed with urea in grassland and barley fields (McTaggart *et al.*, 1997), 63% in lab condition (Pathak and Nedwell, 2001) and 52.6% in winter wheat when fertilized with urea (Bronson and Mosier, 1993).

Conclusion

From the foregoing results of present study, it could be inferred that both LCC based N management along with 10% of DCD registered lower GHG emission. It is also reasonable to suggest that DCD have potential to decrease GHG emission and other wise increase the efficiency of N cycle. They are potentially additional tool to assist agriculture to achieve its economic and environmental goals considerably. Hence, as a simple tool, LCC based N management along with 10% of DCD is found to be the optimal N fertilization strategy for rice, since it gives lower GHG emission besides savings of N as compared to blanket N recommendation. More research is required therefore to move this emerging technology from the current "proof of concept" situation to a practical, cost effective technology on the farm

References

- Bouwman, A.F. 1990. Exchange of greenhouse gases between terrestrial ecosystems and the atmosphere. In: Bouwman AF (ed) Soil and the Greenhouse Effect. John Wiley and Sons, New York, USA, pp: 62–127.
- Bronson, K and A. Mosier. 1993. Nitrous oxide emissions and methane consumption in wheat and corn-cropped systems in Northeastern Colorado. In: Harper, L. A., A. R. Mosier, J. M. Duxbury and D. E. Rolston (eds.), Agricultural ecosystem effects on trace gases and global climate change. Madison. pp: 133–144.
- Di, H.J. and K.C. Cameron. 2004. Effects of the nitrification inhibitor dicyandiamide on potassium, magnesium and calcium leaching in grazed grassland. *Soil use and management.* **20:** 2-7.
- Eichner, M.J. 1990. Nitrous oxide emissions from fertilized soils; Summary of available data. *J. Environ. Qual.* **19:** 272–280
- Hussain, S., Peng, S., Fahad, S., Khaliq, A., Hunag, J., Ciu, K. And Nie, L. 2015. Rice management interventions to mitigate greenhouse gas emission: a review. Environ. Sci. Pollut. Res. 22: 3342–3360.
- Irigoyen,I., Muro, J., Azpilikuefa, M., Aparicio T.P. and C. Lamyfus. 2003.Ammonium oxidation kinetics in the presence of nitrification inhibitors DCD and DMPP at various temperatures. Australian Journal of Soil research. 41:1177-1183.
- Kumar, S.S. and S.K. Malyan. 2016. Nitrification Inhibitors: A perspective tool to mitigate green house gas emission from rice soils. Curr. World Environ. 11(2): 423-428
- Majumdar, D and S. Mitra. 2004. Methane consumption from ambient atmosphere by a typical Ustochrept soil as influenced by urea and two nitrification inhibitors. *Biology and Fertility of Soils*. **39**:140–145.
- Malla, G., Bhatia, A., Pathak, H., Prasad, S., Jain, N. and J. Singh. 2005. Mitigating nitrous oxide and methane emissions from soil in rice-wheat system of the Indo-Gangetic plain with nitrification and urease inhibitors. *Chemosphere*. 58:141–147.
- McTaggart, I., Clayton, H., Parker, J., Swan, L. and K. Smith. 1997. Nitrous oxide emissions from grassland and spring barley, following N fertilizer application with and without nitrification inhibitors. *Biology and Fertility of Soils*. 25: 261–268.

- Mosier, A. R. 1998. Soil processes and global change. *Biology and Fertility of Soils.* **27**: 221–229.
- Mosier, A. R., Duxbury, J. M., Freney, Jr., Heinemeyer, O., Minami, K. 1996. Nitrous oxide emissions from agricultural field, assessment, measurement and mitigation. *Plant and Soil*. **181**: 95–108.
- Pathak, H and D. B. Nedwell. 2001. Nitrous oxide emission from soil with different fertilizers, water levels and nitrification inhibitors. *Water Air and Soil Pollution*. **129:** 217–228.
- Prasad, R and P.J. Power. 1995. Nitrification inhibitors for the agriculture health and environment. *Adv. Agron.* **54:** 233–281.
- Rees, R. M., Baddeley, J. A., Bhogal, A., Ball, B. C., Chadwick, D. R., Machael, M., Lilly, A., Pappa, V. A., Thorman, R. E., Watson, C. A. and J. R. Williams. 2013. Nitrous oxide mitigation in UK agriculture, *Soil Science and Plant Nutrition*. **59:** 3-15.
- Skiba, U., Smith, K.A., and D. Fowler. 1993. Nitrification and denitrification as source of nitric oxide and nitrous oxide. Soil Biology and Biochemistry. 25: 1527–1536.
- Wang, Z., Van Cleemput, O. and L. Baert. 1991. Effect of urease inhibitors on denitrification in soil. Soil Use Manag. 7: 230–233.
- Zacherl, B and A. Amberger. 1990. Effect of Nitrification inhibitors dicyandiamide, nitrapyrin and thiourea on *Nitrosomonas europaea. Fertilizer Research.* 22: 37–44.

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