

#### **RESEARCH ARTICLE**

# Field Efficacy of Certain Biorational Pesticides Against Aphid, *Aphis gossypii* Glover (Hemiptera: Aphididae) on Watermelon, *Citrullus lanatus* Thunb. Matsum and Nakai

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#### ABSTRACT

Aphis gossypii Glover is the major sucking pest in watermelon in early stage of the crop resulting 45-75 % of seedling mortality. In the present study, a field experiment was conducted to evaluate certain biorational pesticides against A. gossypii in watermelon. It was found that a significant lower number of A. gossypii population (4.39 aphid/leaf) was recorded with the treatment of Azadirachta indica oil @ 3% followed by Beauveria bassiana  $(1x10^{8} \text{ cfu spores} @ 8 (g.L^{1})(6.01 \text{ aphid/leaf}) \text{ and spinosad } 45\% \text{ SC } @$ 0.3(mL.L<sup>-1</sup>) (9.16 aphid/leaf), respectively resulting 92.89%, 90.27% and 85.17% reduction of A. gossypii population. Similarly, with the same trend the highest fruit yield (25.50 t ha<sup>-1</sup>) was recorded in the treatment of A. indica followed B. bassiana (24.43 t ha-1), spinosad (23.53 t ha-1) and the lowest in the untreated check (16.30 t ha-1). The increase in fruit yield (56.44%) was recorded in the treatment of A. indica followed B. bassiana, (49.87%) and spinosad (44.17%). A maximum incremental benefit-cost ratio of 1:2.84 was recorded in the treatment of A. indica followed by B. bassiana (1:2.74) and spinosad (1:2.68). From these findings it may be recommended that three rounds of application of A. indica oil @ 3% or B. bassiana (1x108cfu spores @8g /L) or spinosad 45% SC@ 0.3 (mL.L<sup>1</sup> )at 15 days interval for the management of A. gossypii in watermelon

Keywords: Watermelon; Aphis gossypii; Leaf damage; Biorational pesticides

# INTRODUCTION

Watermelon is an important horticultural crop grown for consumption and export purpose. It is a staple food both in fresh and preserved form. It is cultivated throughout the tropical zones of the world (Anonymous, 1988), It is an annual vining plant belonging to the family Cucurbitaceae. The fruit is rich in vitamins A, B1, B2, C, and minerals (Moniruzzaman, 1988). More than 35 varieties/ hybrids like, Arka Manik, PKM 1, Arka Jyoti, Pusa Bedana, Durgapur Kesar, NS707, Namdhari-NS-34, Asahi Yamato and Ice box hybrids (Sugar Baby, Madhuri, Black Magic. Melody, Maxx, Sugar, Super queen, Kalash, Melody, Mahima, Sahana Queen) are grown in India. It is cultivated to the extent of 1.03 lakh ha with an annual production of 25.04 lakh mt ha-1 in India (Anonymous, 2019). It is cultivated in 6,420 ha in Tamil Nadu with annual production of 1.75 mt ha-1 with an average productivity 32 t ha<sup>-1</sup> (Potnuru Santosh Kumar and Kulkarni 2018). It is also extensively cultivated in Tamil Nadu during kharif, Rabi and summer seasons and predominantly in the districts of Villupuram, Namakkal, Ariyalur, Coimbatore and Erode (Chadha, 2013).

Several insect pests attack watermelon at various stages that ravage its cultivation (Anonymous, 2012). The cotton aphid, *Aphis gossypii* was the most destructive pests of watermelon in the early stage of crop growth. They are minute pear-shaped, soft-bodied insects with high reproductive potential and ability to transmit viruses non-persistent manner. They suck sap from tender plants, leading to curling of leaves and distortion, especially when the population is high. They excrete lot of honeydews and sugar-rich substrates that promote the growth of sooty mold (*Capnodium* spp.) on plant parts and leaves lowering their photosynthetic efficiency. The

farmers resort to the application of several rounds of various unrecommended insecticides against *A. gossypii*, which are harmful to human beings and the environment (Anonymous, 1991). Hence, a study was under taken to find effective and eco friendly biorationals on watermelon.

# **MATERIALS AND METHODS**

The field experiment was carried out in a farmer's field at Konur village (11° 12'30" N, 78° 4'41" E"), Namakkal district of Tamil Nadu during summer season 2020 to evaluate field efficacy of certain biorational pesticides against *A. gossypii* in comparison with farmers practice in Randomized Block Design (RBD) with fourteen treatments (Table.1)

#### Table1. List of biorational pesticides tested against A.gossypii in watermelon

#### **Treatments details**

- T<sub>1</sub>- Vitex negundo leaf decoction 5%
- T<sub>2</sub> Azadirachta indica oil 3%
- T<sub>3</sub> Pongamia pinnata -leaf decoction 3%
- $T_{a}$  Ocimum sanctum- leaf decoction 5%
- T<sub>5</sub> Ricinus communis -oil 3%
- T<sub>e</sub> Eucalyptus globulus leaf decoction 3%
- T<sub>7</sub> Beauveria bassiana (1x10<sup>8</sup> cfu spores) -8g.L-1<sup>-1</sup>
- $T_8$  Metarhizium anisopliae-(1x10<sup>8</sup> cfu spores) 8g.L<sup>-1</sup>
- $\rm T_{\rm q}$  Paecilomyces fumosoroseus -(1x10 $^{\rm 8}\, cfu$  spores) 8g.L $^{\rm 1}$
- T<sub>10</sub> Lecanicillium lecanii (1x10<sup>8</sup> cfu spores) 8g.L<sup>-1</sup>
- T<sub>11</sub> Emamectin benzoate 5% SG@0.4g.L <sup>-1</sup>
- T<sub>12</sub> Spinosad 45% SC@ 0.3mL.L<sup>-1</sup>
- T<sub>13</sub> Imidacloprid17.8%SL@0.3mL.L<sup>-1</sup> (Treated check)
- T<sub>14</sub> Untreated check

Three replications were maintained for each treatment. The popular hybrid  $F_1$  (Melody) was sown in protray and 13 days old seedlings were transplanted in the main field at a spacing of 2.5 m x 0.5 m and other recommended package of practices were adopted to raise the crop successfully.

The first spray was given with the onset of pest incidence after recording the pretreatment count. The subsequent sprays were repeated after 15 days using a high volume sprayer with required concentrations. The posttreatment counts on pests were recorded on 1, 3, 7, 14 days after each application. Ten plants were selected randomly from each replication in a plant and *A. gossypii* populations were recorded from three leaves in infested branches (one from the top of the terminal twig) with the unopened leaves and two from opened leaves, and the mean number was calculated per leaf and expressed as number/leaf. The efficacy

of the treatments was assessed based on the category by using a number of population-level of aphids as per Sikha Deka *et al.* (2016)

| Aphid population /leaf | Category    |
|------------------------|-------------|
| <1. No. of aphid       | Negligible  |
| 1-10. No. of aphid     | Low         |
| 11-30. No. of aphid    | Moderate    |
| 31-40. No. of aphid    | Severe      |
| >50. No. of aphid      | Very Severe |

Watermelon fruits were harvested from all three replications and pooled to arrive fruit yield. The yield data were also recorded in untreated plots and the increase in yield and income from each treatment over untreated checks were worked out. Accordingly, the incremental cost-benefit ratio was worked out for all the treatments by using the following formula ICBR = Gross income / (total cost of cultivation + cost of plant protection) as adopted by Akila and Sundara Babu (1994) where the cost of plant protection = cost of insecticide + labour charges for spraying.

#### Statistical analysis

The data generated from the field experiments were analyzed for ANOVA. The data on insect population were transformed into square root transformation and analyzed using SPSS (version 22) (IBM crop Released 2013) software to identify the most effective treatments and their means were compared by significant difference at p<0.05 ANOVA followed by Tukey's Honest Significant Difference test.

### **RESULTS AND DISCUSSION**

The results revealed that all the biorational pesticides were effective in reducing *A. gossypii* infestation. The data on *A. gossypii* population per leaf percent reduction of the population over untreated check, increase in yield and ICBR due to biorationals are furnished in Table 1 and Fig 1.

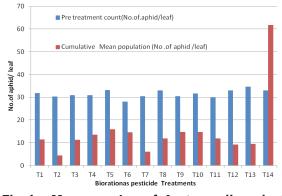


Fig 1. Mean number of *A. gossypii* against biorational pesticides

The lowest population of *A. gossypii* in the treatment of *A. indica* (4.39/leaf) was recorded followed by *B. bassiana* (6.01 /leaf), spinosad (9.16/leaf) and the highest number of *A. gossypii* was observed in the untreated check (61.79/leaf). Further, the order of efficacy of the treatments was imidacloprid (Treated check) (9.57 aphid/leaf) < *P. pinnata* (11.36 aphid/leaf) < *V. negundo* (11.38 aphid/leaf) < emamectin benzoate (11.86 aphid/leaf) < *M. anisopliae* (11.95 aphid/leaf) < *O. sanctum* (13.59 aphid/leaf) < *E. globulus* (14.64 aphid/leaf) < *L. lecanii* (14.74 aphid/leaf) < *P. fumosoroseus* (14.75 aphid /leaf) < *R. communis* (15.93 aphid /leaf).

The observation on *A.* gossypii population density in the treatment of *A.* indica was recorded as the lowest category followed by

*B.* bassiana, spinosad and imidacloprid. The moderate category was recorded in *P. pinnata*, *V. negundo*, emamectin benzoate, *M. anisopliae*, *O. sanctum*, *E. globulus*, *L. lecanii*, *P. fumosoroseus*, *R. communis* and the very severe category were in the untreated control respectively (Table 2).

Similarly, the reduction of *A. gossypii* population was high in *A. indica* (92.89%) followed by *B. bassiana* (90.27%) and spinosad (85.17%). Further, the order of the reduction over treated check (Imidacloprid) was 84.51%, followed by *P. pinnata* (81.61%) > *V. negundo* (81.58%) >emamectin benzoate (80.80%) > *M. anisopliae* (80.66%) > *O. sanctum* (78.05%) > *E. globulus* (76.30%) > *L. lecanii* (76.14%) > *P. fumosoroseus* (76.12%) > *R. communis* (74.21%).

\*NS - Non significant

|                       | Treatments details  |              | ***Cumulative Mean<br>Number of aphids/<br>top three Leaves<br>-(X±SE)*** | Aphid<br>population<br>(Category) | Per cent<br>reduction over<br>untreated check | (+ ha_1)***               | Per cent<br>increasing yield<br>over untreated<br>check | ICBR   |
|-----------------------|---|--------------|---|-----------------------------------|---|---------------------------|---|--------|
| T <sub>1</sub>        | Vitex negundo - leaf decoction 5%   | 31.88 (5.65) | 11.38 (3.37)±1.40 <sup>abc</sup>  | Moderate                          | 81.58(9.03)                                   | 19.93(4.46) <sup>fe</sup> | 22.26(4.72) <sup>f</sup>                                | 1:2.40 |
| $T_2$                 | Azadirachta indica - oil 3%   | 30.30 (5.50) | 4.39 (2.09)±0.63 °  | Low                               | 92.89(9.64)                                   | 25.5(5.05) <sup>a</sup>   | 56.44(7.51) <sup>a</sup>                                | 1:2.84 |
| $T_{_3}$              | Pongamia pinnata -leaf decoction 3%   | 30.95 (5.56) | 11.3 (3.37)±1.11 bc   | Moderate                          | 81.61(9.03)                                   | 22.9(4.79) <sup>bc</sup>  | 21.93(4.68) <sup>f</sup>                                | 1:2.31 |
| $T_4$                 | Ocimum sanctum- leaf decoction 5%   | 30.88 (5.56) | 13.5 (3.69)±1.23 bc   | Moderate                          | 78.00(8.83)                                   | 19.6(4.43) <sup>gfe</sup> | 20.24(4.5) <sup>fg</sup>                                | 1:2.48 |
| $T_5$                 | Ricinus communis -oil 3%  | 33.16 (5.76) | 15.93(3.99)±1.34 °  | Moderate                          | 74.21(8.61)                                   | 23.03(4.8) <sup>bc</sup>  | 41.28(6.42)°  | 1:2.59 |
| Т <sub>6</sub>        | Eucalyptus globulus - leaf<br>decoction 3%                                      | 28.04 (5.30) | 14.64(3.83)±1.38 °  | Moderate                          | 76.30(8.73)                                   | 21.53(4.64) <sup>cd</sup> | 31.9(5.65) <sup>d</sup>                                 | 1:2.46 |
| <b>T</b> <sub>7</sub> | Beauveria bassiana (1x10 <sup>s</sup> cfu<br>spores) -8g.L <sup>-1</sup>        | 30.53 (5.53) | 6.01(2.45)±0.80 bc  | Low                               | 90.27(9.50)                                   | 24.43(4.94) <sup>ab</sup> | 49.87(7.06) <sup>b</sup>                                | 1:2.74 |
| T <sub>8</sub>        | Metarhizium anisopliae-<br>(1x10 <sup>s</sup> cfu spores) 8g.L <sup>.1</sup>    | 32.95 (5.74) | 11.95(3.46)±1.26 bc   | Moderate                          | 80.66(8.98)                                   | 18.5(4.3) <sup>gf</sup>   | 13.49(3.67) <sup>h</sup>                                | 1:2.17 |
| T <sub>9</sub>        | Paecilomyces fumosoroseus<br>-(1x10 <sup>8</sup> cfu spores) 8g.L- <sup>1</sup> | 30.54 (5.53) | 14.75 (3.84)±0.95 °   | Moderate                          | 76.12(8.72)                                   | 18.17(4.26) <sup>g</sup>  | 11.41(3.38)   | 1:2.20 |
| T <sub>10</sub>       | Lecanicillium lecanii (1x10 <sup>s</sup> cfu<br>spores) 8g.L <sup>.1</sup>      | 31.64 (5.62) | 14.74 (3.84)±1.19 °   | Moderate                          | 76.14(8.73)                                   | 19.5(4.42) <sup>gfe</sup> | 19.63(4.43) <sup>g</sup>                                | 1:2.34 |
| T <sub>11</sub>       | Emamectin benzoate 5%<br>SG@0.4g.L <sup>1</sup>                                 | 30.09 (5.49) | 11.86 (3.44)±1.28 bc  | Moderate                          | 80.80(8.99)                                   | 20.37(4.51) <sup>ed</sup> | 25.15(5.01) <sup>e</sup>                                | 1:2.40 |
| T <sub>12</sub>       | Spinosad 45% SC@ 0.3m.L-1   | 32.96 (5.74) | 9.16 (3.03)±0.87 <sup>abc</sup>   | Low                               | 85.17(9.23)                                   | 23.53(4.85) <sup>b</sup>  | 44.17(6.65) <sup>c</sup>                                | 1:2.68 |
| T <sub>13</sub>       | Imidacloprid17.8%SL@0.3ml.L-<br>(Treated check)                                 | 34.72 (5.89) | 9.57 (3.09)±1.11 <sup>abc</sup>   | Low                               | 84.51(9.19)                                   | 23.50 (4.85) <sup>b</sup> | 44.04 (6.64) <sup>c</sup>                               | 1:2.68 |
| T <sub>14</sub>       | Untreated check   | 33.06 (5.71) | 61.79 (7.86)±3.40 <sup>d</sup>  | Sever                             | -   | 16.3(4.04) <sup>h</sup>   | -   | -      |
| F                     | -   | -            | 28.80   | -                                 | -   | 80.44                     | 1963.98   | -      |
| Ρ                     | -   | *-NS-        | **<0.000  | -                                 | -   | **< 0.000                 | **< 0.000   | -      |
| SD                    | -   | -            | 13.85   | -                                 | -   | 2.69                      | 16.33   | -      |
| SE                    | -   | -            | 3.70  | -                                 | -   | 0.72                      | 04.37   | -      |

| Table 2. | <b>Biorational pest</b> | icides against Aph | nis gossypii on | watermelon |
|----------|-------------------------|--------------------|-----------------|------------|
|----------|-------------------------|--------------------|-----------------|------------|

PTC- Pre treatment count DAS-Days after spraying, F= F value of Tukeys Test P=Statistical significant.

SE= Standard error \*\*Highly significant SD= Standard deviation ICBR-Incremental cost benefit ratio \*\*\*\* Sale price of watermelon was Rs.5.00 per \*\*\* Each value is the mean of three replications.

Figures in parentheses are square root transformed values.

In a column, means followed by common letter(s) is /are not significantly different by Tukey HSDs test at P=0.05%

A significantly highest fruit yield (25.50 t ha<sup>-1</sup>) was recorded in the treatment of *A. indica* followed by in *B. bassiana*, (24.43 t ha<sup>-1</sup>), spinosad (23.53 t ha<sup>-1</sup>) and lowest in untreated check (16.30 t ha<sup>-1</sup>). Further, the orders of fruit yields were in imidacloprid (23.50 t ha<sup>-1</sup>) >*R. communis* (23.03 t ha<sup>-1</sup>) > *P. pinnata* (22.90 t ha<sup>-1</sup>) > *E. globulus* (21.53 t ha<sup>-1</sup>) > *emamectin* benzoate (20.37 t ha<sup>-1</sup>) > *V. negundo* (19.93 t ha<sup>-1</sup>) > *O. sanctum* (19.60 t ha<sup>-1</sup>) > *L. lecanii*, (19.50 t ha<sup>-1</sup>) > *M. anisopliae* (18.50 t ha<sup>-1</sup>) > *P. fumosoroseus* (18.17 t ha<sup>-1</sup>) (treated check) and (untreated check) (16.30 t ha<sup>-1</sup>) respectively.

The increase in fruit yield (56.44%) was recorded in the treatment of *A. indica* followed by *B. bassiana*, (49.87%) and spinosad (44.17%). Further, the order of increase in fruit yield over treated check was in imidacloprid (44.04%) >*R. communis* (41.28%)>*E.* globulus (31.90%) > emamectin benzoate (25.15%) > *V. negundo* (22.26%) > *P. pinnata* (21.93%) >*O.* sanctum (20.24%)> *L. lecanii* (19.63%) > *M.* anisopliae (13.49%) > *P. fumosoroseus* (11.41%).

A maximum ICBR of 1:2.84 was recorded in the treatment of *A. indica* followed by in *B. bassiana*, (1:2.74) and spinosad (1:2.68) followed by imidacloprid (1:2.68) > *R. communis* (1:2.59). > 0. sanctum (1:2.48) > *E. globulus* (1:2.46) > *V. negundo* (1:2.40) > emamectin benzoate (1:2.40) > *L. lecanii* (1:2.34) > *P. pinnata* (1:2.31), *P. fumosoroseus* (1:2.20) > *M. anisopliae* (1:2.17).

Nur et al. (2020) reported that biorational insecticides are found to have repellent, antifeedant and mortality effects on herbivore insects and affect their reproduction, growth and development thus reducing their population and infestation on Lablab purpureus. Our finding on the watermelon crop agrees with Sedlacek and Townsend (1990) that the heavy infestations by aphids are common on cucurbitaceous and solanaceous crops. In our study, the biorational insecticides tested were found to have significantly lowered the infestation of A. gossypii. These findings are in accordance with Amin et al. (2017) and Mohammad et al. (2018) who observed the reduction of A. gossypii population and infestation in bitter gourd with sequential application of bio-pesticide, botanical insecticides.

Lowery et al (1993) found that neem oil 3% found to be very effective against *A. gossypii* under laboratory condition. The present study is in accordance with the findings of Sardana et al. (2004). Khalequzzaman and Nahar (2008) found that *A. indica* was more toxic than imidacloprid, malathion, and carbosulfan to control four aphid species namely *Aphis craccivora*, *A. gossypii*, *Myzus persicae* and *Lipaphis erysimi* on bean, brinjal, potato and cauliflower plants respectively. Mandal et

al. (2006) evaluated various neem based products and found that neem oil (3%) was effective against *A. gossypii* in okra which is supporting our findings. Dimetry et al. (2013) found that Nimbecidine @ 0.03% + microbial product of *L. lecanii* spores @1×10<sup>9</sup> were effective against aphid, *M. persicae* in cucumber which is (Mahesar et.al.,2011; Naeem et.al.,2012) supporting our findings.

Muhammad et al. (2013) confirmed the effectiveness of *B. bassiana*  $1 \times 10^6$ ,  $1 \times 10^7$  and  $1 \times 10^8$  spores/mL on *A. gossypii* and found the uppermost concentration ( $1 \times 10^8$  spores/mL) proved to give maximum control within short period of time. Similar observations were made by Arun et al. (2018) and they reported that the entomopathogenic fungi, *B. bassiana* were found to be effective against *L.erysimi*. These findings are in line with our observations. Khadija Javed et al. (2019) confirmed that maximum aphid mortality was observed with the treatment of *L. lecanii* and *B. bassiana* under lab conditions correlates with our results in watermelon even under field condition also.

Ahmed et al. (2016) found the role of *B.bassiana* against the adults of cabbage aphid, *Brevicoryne* brassicae under laboratory conditions at three tested concentrations ( $10^5$ ,  $10^6$  and  $10^7$  spores/mL). After 7 days of application, the resulting mortality percentages were 37.77%, 60.00% and 73.33% respectively under field conditions. This finding is supported by our observation. Ravi and Nakat (2017) found that the combination of Entomopathogenic fungi as *L lecanii* 1.15 % WP + *M. anisopliae* 1.15 % WP was the most effective treatment as compared to standard check dimethoate suppressing of *A.gossypii* population in Okra. Similar findings by Arun Janu et al., (2018) were found in *A. gossypii* which correlate with the present investigation.

Rosalind et al. (1995) showed that strain of B. bassiana (1×10<sup>15</sup>spores/ mL) reduced pea aphid, Acyrthosiphon pisum population up to 97.9 % under field conditions. Khan et al. (2012) also proved the same results showing 80% mortality caused by the filtrate application of *B. bassiana* while 57% mortality caused by conidia on 6<sup>th</sup> day of application. The present findings are in close agreement with Singh et al. (2008) who reported that L. lecanii @ 10<sup>8</sup> spores(mL<sup>-1</sup>) dose was effective in controlling the aphid population to 75.79 %. While, Kadam et al. (2008) showed that the L. lecanii @ 6 × 10<sup>5</sup> (cfu .m.L<sup>-1</sup>) 0.3 % had reduced the initial population of M.persicae in mustard. Palande and Pokharkar (2005) reported that the biological activity of V. lecanii against B brassicae with mortality from 16.3 to 93.3 % by the concentrations of  $1 \times 10^3$  to  $1 \times 10^9$  cfu/ mL. are in line with our investigation.

Similarly, Poprawski et al. (1999) also reported that the B. bassiana based myco insecticide was effective against brown citrus aphid, Toxoptera citricida and observed rapid kill of 94.4% and 79.8% with 5  $\times$  10<sup>13</sup> and 2.5  $\times$  10<sup>13</sup> conidia ha<sup>-1</sup> respectively. Neelam et al. (2003) tested the L. lecanii at the concentration of 10<sup>6</sup>, 10<sup>7</sup>,10<sup>8</sup> conidia mL<sup>-1</sup> against L. erysimi and reported the highest mortality of 80% 96 hrs of treatment at the concentration of 10<sup>8</sup> conidia mL<sup>-1</sup>. Nirmala et al. (2006) and Asi et al. (2009) found that the, L. lecanii, M. anisopliae and P. fumosoroseus can effectively control aphid. Furthermore, Vu et al. (2007) reported that among the fungi tested, L. lecanii, P.fumosoroseus B. bassiana, M.anisopliae, Cordyceps scarabaeicola and Nomuraea rileyi, performed as the best in controlling A. gossypii.

Suganthy and Sakthivel (2012) found that the maximum population reduction was observed in neem oil 3% treatment (2.6 aphid/ plant), followed by NSKE (2.8 aphid/ plant) and V. negundo (3.1 aphid/ plant) in Solanum nigrum. The results were following the findings of Uthamasamy and Gajendran (1992), Belmain et al. (2001) who confirmed that V.negundo @ 2% recorded the highest mortality of A. gosspii with strong repellence from (77.11%). Kulat et al. (1997) have also indicated that pongam leaf extract was highly toxic to the A. gossypii in okra. Das et al. (2008) found that spinosad -treated plants provided a higher yield of lab lab. Among the botanicals neem oil showed a significant reduction of aphid population and consequently gave better yield (1.22 kg/plot) which are in conformity with our investigation Patel et al. (2015) reported that emamectin benzoate @ 10 g.a.i ha-1 was found to be most effective as it recorded the lowest infestation of all the recorded sucking pest of brinjal. Nur et al. (2020) found that the treatments viz, spinosad, emamectin benzoate and neem oil have exerted the lowest aphid population and resulted in significantly higher marketable yield than that of the control on Lablab purpureus. These records are comparable with our investigation in watermelon. The effectiveness of R. communis oil in our experiment is also comparable with the results of Arya et al. (2014) in the mustard aphid.

# CONCLUSION

It is inferred that among the thirteen bio rational pesticides evaluated for their efficacy against *A. gossypii* in watermelon three rounds of application of *A. indica* oil @ 3% or *B. bassiana* (1x10<sup>8</sup>cfu spores @ 8g/Lit) or spinosad 45% SC @ 0.3mL/Lit at 15 days interval could be adopted for the management of *A. gossypii* in watermelon

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### **Ethics statement**

No specific permits were required for the described field studies because no human or animal subjects were involved in this research.

#### **Consent for publication**

All the authors agreed to publish the content.

#### **Competing interests**

There were no conflict of interest in the publication of this content

#### Data availability

All the data of this manuscript are included in the MS. No separate external data source is required. If anything is required from the MS, certainly, this will be extended by communicating with the corresponding author through corresponding official mail: csankarento@gmail.com

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