

# RESEARCH ARTICLE Optimization of Cultural Conditions for the Antimetabolites Production by *Streptomyces aureus* strain BG03

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# ABSTRACT

| · | ust, 2018<br>tember, 2018<br>tember, 2018 | The growth of <i>Streptomyces aureus</i> strain BG03 was the best on Ken Knight`s agar medium. Among the different media tested, Crawford`s media was the best for the production of bioactive compounds by S. aureus strain BG03. The culture filtrate of <i>S. aureus</i> strain BG03 extracted from Crawford`s broth amended with starch as a carbon source, peptone as a nitrogen source, temperature of $35$ °C and pH 7.5 recorded the highest growth inhibition and inhibition zone against <i>Rhizoctonia bataticola</i> causing dry root rot in urdbean. Therefore the above mentioned parameters were optimum for the maximum production of bioactive compounds by <i>S. aureus</i> strain BG03. The crude metabolites extracted from S. aureus strain BG03 was found to be promising in reducing the mycelial growth of <i>R. bataticola</i> . Gas Chromatography-Mass Spectrophotometry (GC-MS), analysis of crude metabolites of <i>S. aureus</i> strain BG03 revealed the presence of seven compounds having antifungal activity. |
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## Keywords: Streptomyces aureus, Optimization, Cultural conditions, Antimetabolite production,

Streptomyces is the largest genus of the actinomycetes which are Gram-positive, aerobic, non-motile with a filamentous form that resembles fungi. *Streptomyces* have relatively large genomes of approximately 8–9 Mbp in size with a high GC content of more than 70 per cent (Singh *et al.*, 2012). *Streptomyces* are prolific producers of wide variety of bioactive compounds which are found to have antifungal (Lim *et al.*, 2015), antibacterial, antiviral (Abd and Hatem, 2013), immunosuppressive, and antioxidant properties (Wang *et al.*, 2008). They account for nearly 60 per cent of the production of agriculturally important antibiotics (Liu *et al.*, 2013).

The production of secondary metabolites from the genus *Streptomyces* can be influenced by optimization of the nutritional requirements and cultural conditions. These conditions play an important role in the production of these secondary metabolites (Khattab *et al.*, 2016). Antibiotic biosynthesis is a specific property of microorganisms which depends greatly on culture conditions. Improvement in the growth and antibiotic production can be carried out by manipulating the nutritional and physical parameters of the culturing conditions (Gao *et al.*, 2009). The present work was carried out with a view to investigate the effect of different media, carbon and nitrogen sources, on bioactive compound production by the *S. aureus* strain BG03. The influence of pH and temperature on bioactive compound production was also studied.

# **MATERIAL AND METHODS**

# Cultural characteristics of S. aureus strain BG03

Cultural characteristics *viz.,*growth pattern, color of aerial mycelium, color of substrate mycelium, and surface characteristics of the S. *aureus* strain BG03 were recorded on different media *viz.,*Ken Knight`s, Crawford`s, Potato dextrose, Yeast extract malt extract, Nutrient, Starch casein hydrolysate, starch agar, AF/ MS, and Tryptone soya agar medium.

# Optimization of cultural and nutritional conditions for the production of bioactive compounds

Streptomyces aureus strain BG03 was inoculated in different broth and incubated for 10 d at 30 °C on a rotary shaker (150 rpm) to find out the broth which produces maximum bioactive compound. The media used were Knight`s, Crawford`s, Potato dextrose, Yeast extract malt extract, Nutrient, Starch casein hydrolysate, AF/ MS, and Tryptone. After incubation actively grown culture of 100 ml quantity from each broth was centrifuged at 10,000 rpm at 4°C for 10 min and the supernatant containing, culture filtrate of respective broth was

obtained. From that culture filtrate 400µl of each broth were taken aseptically and the antifungal activity was measured using agar well diffusion assay.

Nitrogen source of Crawford`s broth (Yeast extract) was substituted with potassium nitrate, peptone, beef extract and malt extract inoculated with S. *aureus* strain BG03 and incubated for 10 d at 30°C on a rotary shaker (150 rpm). Culture filtrate of each nitrogen source broth was used to identify antifungal activity against *R. bataicola* by agar well diffusion assay.

The S. *aureus* strain BG03 was inoculated on Crawford`s broth (pH 7.0) at various levels of pH 6, 7.5, 8 and 9 using a phosphate buffer and incubated for 10 d at 30°C on a rotary shaker (150 rpm) to optimize the pH for maximum bioactive compound production in broth medium. Culture filtrate of each pH source broth was used to identify antifungal activity against *R. bataicola* by agar well diffusion assay.

The S. aureus strain BG03 was inoculated in Crawford's broth and incubated at different temperatures 20,25, 30, 35 and 40°C for determining the optimum temperature for maximum bioactive production. After the incubation period the culture filtrate of broth at above mentioned temperature was prepared and tested for antifungal activity against *R. bataicola* by agar well diffusion assay.

## Efficacy of crude metabolites against Rhizoctonia bataticola

The S. aureus strain BG03 was inoculated into Crawford's broth in a 250 ml conical flask, and the culture was incubated at 28 °C for 48 h on a shaker at 120 rpm. At the end of incubation period the culture filtrate was collected by centrifugation at 10,000 rpm for 15 min and the supernatants were collected. Antimicrobial compound was partially purified from the culture filtrate by solvent extraction method. Ethyl acetate was added to the culture filtrate in the ratio of 1:1 (V/V) and shaken vigorously for 1 h for complete extraction. The ethyl acetate phase that contains bioactive metabolites was separated from the aqueous phase by using separating funnel. The ethyl acetate fraction was evaporated to dryness in a water bath and residue obtained was used to determine antifungal activities against *R. bataticola* by agar well diffusion method and effectiveness was measured by zone of inhibition.

## **Characterization of antibiotics**

The crude metabolites from the S. *aureus* strain BG03 was analyzed through Gas Chromatography-Mass spectrometry (GCMS) at Agricultural College and Research Institute, Madurai. GC-MS analysis was carried out on a Shimadzu GCMS-QP2010 Ultra system. The injector temperature was 280 °C. The samples were injected in the split mode with split ratio 1/25. Injection volume was 1  $\mu$ L. A capillary column Rtx-5MS (5% Diphenyl-95% Dimethyl Polysiloxane), 30 m x 0.25 mm x 0.25  $\mu$ m, was used. Carrier gas was helium with constant flow of 1.00 ml min-1. The oven temperature was as follows: initial temperature of 60 °C, held for 2 min, increased to 10 °C min-1 up to 260 °C and held for 10 min. The MS ionization potential was 70 eV, and the temperatures were as follows: interface 260 °C, lon source 280 °C. Mass scan range 40-550.

# **RESULTS AND DISCUSSION**

The actinobacterial isolate S. *aureus* strain BG03 was grown on different solid media *viz.*,Crawford`s agar, Ken Knight`s agar, Potato dextrose agar, Yeast extract malt extract agar, Nutrient agar, Starch casein hydrolysate agar, Starch agar, AF/MS agar, Oat meal agar, Tryptone soya agar to determine the best media for its growth.

| Madin                           | Cultural characterization |                 |                    |                           |  |  |
|---------------------------------|---------------------------|-----------------|--------------------|---------------------------|--|--|
| Medium                          | Growth                    | Aerial mycelium | Substrate mycelium | Surface                   |  |  |
| Crawford's agar                 | Good                      | White           | Pale yellow        | Slight powdery            |  |  |
| Ken Knight`s agar               | Excellent                 | Whitish yellow  | Pale yellow        | Powdery                   |  |  |
| Potato dextrose agar            | Moderate                  | White           | Pale yellow        | Slight powdery            |  |  |
| Yeast extract malt extract agar | Moderate                  | Pure white      | Pale yellow        | Slimy                     |  |  |
| Nutrient agar                   | No growth                 | -               | -                  | -                         |  |  |
| Starch casein hydrolysate agar  | Excellent                 | White           | Pale yellow        | Slimy                     |  |  |
| Starch agar                     | Excellent                 | White           | Pale yellow        | Slight powdery with Slimy |  |  |
| AF/ MS agar                     | No growth                 | -               | -                  | -                         |  |  |
| Tryptone soya agar              | Good                      | Creamy white    | Pale yellow        | Slimy                     |  |  |
| Oat meal agar                   | No growth                 | -               | -                  | -                         |  |  |

The result indicated that the growth of S. *aureus* strain BG03 was excellent in Ken knight's agar, starch casein hydrolysate and starch agar medium. The colour of the aerial mycelium was white to creamy white and the substrate mycelium was pale yellow in colour and the colony showed powdery to slimy texture. The growth of S. *aureus* strain BG03 was found to be good in Tryptone soya agar, Crawford's agar and it was moderate in Potato dextrose agar, Yeast extract malt extract agar. On oat meal agar and nutrient agar medium no growth of S. *aureus* BG03 was observed (Table 1). Yasmeen *et al.* (2016) found that *Streptomyces* sp. strain VJSY-2 exhibited excellent growth on YMD agar medium and S.*carpaticus* strain VJSY-3 exhibited good growth on tryptone yeast extract agar (ISP-1), and tyrosine agar.

#### Optimization of cultural condition for production of bioactive compounds by S. aureus strain BG03

The actinobacteria S. *aureus* strain BG03 was grown in different medium for 10 d to determine the best medium for the maximum production of bioactive compounds. The culture filtrate of S. *aureus* train BG03 from each culture broth was extracted and it was used for assessing the antifungal activity against the growth of *R. bataticola*. The result showed that S. *aureus* strain BG03 had the highest antifungal activity in Crawford`s broth which recorded 49.26 percent growth inhibition and 40.33 mm inhibition zone. It was followed by starch casein broth with 38.88 per cent and 30.00 mm growth inhibition of *R. bataticola* and inhibition zone respectively. The lowest inhibition of 2.96 per cent and inhibition zone of 0.0 mm was recorded by nutrient broth. Hence, the Crawford`s broth which showed highest growth inhibition of *R. bataticola* was suitable media for bioactive metabolite production (Table 2)

| Medium                     | Mycelial growth of pathogen (mm) | Per cent reduction over<br>control | Inhibition zone (mm)* |
|----------------------------|----------------------------------|------------------------------------|-----------------------|
| Ken Knight`s               | 61.33                            | 31.85° (34.35)                     | 10.00 <sup>d</sup>    |
| Crawford`s                 | 45.66                            | 49.26 <sup>a</sup> (44.57)         | 40.33ª                |
| Potato dextrose            | 78.33                            | 12.96 <sup>f</sup> (21.10)         | $00.20^{\mathrm{f}}$  |
| Yeast extract malt extract | 69.00                            | 23.33° (28.88)                     | 10.46°                |
| Nutrient                   | 87.33                            | 2.96 <sup>g</sup> (09.90)          | $0.00^{h}$            |
| Starch casein              | 55.00                            | 38.88 <sup>b</sup> (38.57)         | 30.00 <sup>b</sup>    |
| AF/ MS                     | 68.66                            | 23.71° (29.13)                     | 00.76 <sup>e</sup>    |
| Tryptone                   | 65.00                            | 27.77 <sup>d</sup> (31.80)         | 00.04 <sup>g</sup>    |
| Control                    | 90.00                            | $0.00^{h}(0.64)$                   | $00.00^{h}$           |
| CD(p=0.05)                 |                                  | 0.10                               |                       |

| Table 2. Effect of different media on the antifungal activity of S. aureus strain BG03 culture filtrate against |
|---|
| R. bataticola.  |

\*Values are mean of three replications

Values in parentheses are arcsine transformed values

In a column, means followed by a common letter are not significantly different at the 5% level by DMRT

In this investigation S. *aureus* strain BG03 grown in Crawford`s broth exerted maximum inhibition of *R. bataticola* with highest inhibition zone. This indicated that Crawford`s broth favour the highest production antibiotics of S.*aureus* when compared to other media. Starch casein broth ranked next in enhancing the production of bioactive compounds. Singh *et al.* (2017) found that starch casein broth was the best medium for the antibiotic production by S. *chilikensis* strain ACITM-1. *Streptomyces* sp. grown in yeast malt medium showed the maximum growth inhibition of *M. phaseolina* (Verma *et al.*, 2018).

The Crawford`s broth was amended with various carbon sources *viz.*, fructose, sucrose, starch and maltose to determine the best carbon source for bioactive compound production. Among these, culture filtrate of *S. aureus* strain BG03 grown in media amended with starch was found to be significantly superior in inhibiting the growth of *R. bataticola* with 46.30 per cent growth inhibition and 20.10 mm inhibition zone. It was followed by sucrose which exhibited growth inhibition of 33.17 per cent. The metabolites production by *S. aureus* strain BG03 was low in fructose amended medium as it inhibited the mycelial growth of *R. bataticola* only to the extent of 27.91 per cent (Table 3).

Carbohydrate viz., glycerol, maltose, mannose, sucrose and xylose are reported to interfere with production of metabolites (Shobha *et al.*, 2014). In this study among the different carbon source, starch was found to be the best for the production of bioactive compound by S. *aureus* strain BG03 this findings was in concordance

with Osman *et al.* (2011) who found that starch was the best carbon source for antibiotic production. Similar results were also reported by El-Nasser *et al.* (2010) and Singh *et al.* (2017).

The growth and antimetabolite production is greatly influenced by the nature and type of nitrogen source supplemented in the Crawford`s broth. The antifungal activity of S. *aureus* strain BG03 was found to be high 45.93 per cent growth inhibition and 20.10 mm inhibition zone in medium amended with peptone. The culture filtrate of potassium nitrate incorporated medium recorded the lowest growth inhibition (10.34 %) against *R. bataticola* (Table 3).

| Table 3. Effect of nutritional conditions on the antifungal activity of S. aureus strain BG03 culture filtrate |
|--|
| against Rhizoctonia bataticola.  |

| Nutritional |   |                                       |                          |                   |   |                                       |                             |  |  |
|-------------|---|---------------------------------------|--------------------------|-------------------|---|---------------------------------------|-----------------------------|--|--|
|             | Carbon                                    |                                       |                          |                   | Nitrogen                                  |                                       |                             |  |  |
| Source      | Mycelial<br>growth of<br>pathogen<br>(mm) | Per cent<br>reduction<br>over control | Inhibition<br>zone (mm)* | Source            | Mycelial<br>growth of<br>pathogen<br>(mm) | Per cent<br>reduction<br>over control | Inhibition<br>zone<br>(mm)* |  |  |
| Fructose    | 65.33                                     | 27.41 <sup>d</sup> (33.96)            | 0.23 <sup>d</sup>        | Potassium nitrate | 79.66                                     | 10.34 <sup>d</sup> (18.75)            | 0.26 <sup>d</sup>           |  |  |
| Maltose     | 63.83                                     | 29.07°(28.77)                         | 0.33°                    | Peptone           | 48.66                                     | 45.93 <sup>a</sup> (42.66)            | 20.10 <sup>a</sup>          |  |  |
| Starch      | 48.33                                     | 46.30 <sup>a</sup> (25.23)            | 20.10 <sup>a</sup>       | Beef extract      | 78.66                                     | 12.60 <sup>b</sup> (20.79             | 10.80 <sup>b</sup>          |  |  |
| Sucrose     | 59.66                                     | 33.71°(31.62)                         | 0.60 <sup>b</sup>        | Malt extract      | 87.33                                     | 02.90° (09.80)                        | 0.90°                       |  |  |
| Control     | 90.00                                     | 00.00 <sup>d</sup> (09.97)            | $0.00^{d}$               | Control           | 90.00                                     | 00.00° (00.44)                        | 0.00 <sup>e</sup>           |  |  |
| CD (p=0.05) |   | 0.12                                  |                          | CD (p=0.05)       |   | 0.05                                  |                             |  |  |

\*Values are mean of three replications

Values in parentheses are arcsine transformed values

In a column, means followed by a common letter are not significantly different at the 5% level by DMRT.

It is evident from the present study *S. aureus* strain BG03 showed the highest antifungal action against *R. bataticola* when it was grown in medium amended with peptone as a nitrogen source. This was attributed to the addition of peptone in the medium enhanced that production of bioactive compound, therefore peptone was found to be good nitrogen source for antibiotic production. This was in agreement with findings of Bundale *et al.* (2015) who found that medium amended with peptone resulted in maximum bioactive metabolite production in *S. spectabilis*. Singh *et al.* (2017) reported that peptone and potassium nitrate were best the nitrogen source for the highest antifungal activity and biomass production for *S. chilikensis* strain ACITM-1.

pH of the culture medium affects not only growth of the *Streptomyces* but also production of the antibiotics. The result on pH of the medium revealed that *S. aureus* strain BG03 showed the maximum antifungal activity against *R. bataticola* at pH 7.5. At this pH, *S. aureus* strain BG03 recorded the highest growth inhibition 48.74 per cent with the inhibition zone of 20.00 mm. Growing of *S. aureus* at pH 6.0 and 9.0 was not suitable for antimetabolite production as indicated by the lowest growth inhibition of *R. bataticola* at these pH (Table 4).

| Table 4. Effect of physical conditions on the antifunga | I activity of S. aureus strain BG03 culture filtrate |
|---|--|
| against Rhizoctonia bataticola.                         |  |

|             |                          |                                  | Physical                           |             |                          |                                    |  |  |
|-------------|--------------------------|----------------------------------|------------------------------------|-------------|--------------------------|------------------------------------|--|--|
|             |                          | P <sup>H</sup>                   |                                    |             | Temperature (° c)        |                                    |  |  |
| Level       | Inhibition<br>zone (mm)* | Mycelial growth of pathogen (mm) | Per cent reduction<br>over control | Level       | Inhibition<br>zone (mm)* | Per cent reduction<br>over control |  |  |
| 6.0         | 0.36 <sup>b</sup>        | 87.66                            | 12.60 <sup>b</sup> (20.79)         | 20          | 10.40 <sup>d</sup>       | 35.68° (36.67)                     |  |  |
| 7.5         | 20.06ª                   | 80.33                            | 48.74ª (44.27)                     | 25          | 10.76°                   | 14.45° (22.34)                     |  |  |
| 8.0         | 0.03°                    | 87.83                            | 22.90° (28.59)                     | 30          | 20.10 <sup>b</sup>       | 40.74 <sup>b</sup> (39.66)         |  |  |
| 9.0         | $0.00^{d}$               | 88.5                             | 11.60 <sup>d</sup> (19.91)         | 35          | 20.60ª                   | 46.90 <sup>a</sup> (43.22)         |  |  |
| Control     | $0.00^{d}$               | 90.00                            | $0.00^{d} (0.44)$                  | 40          | 0.23°                    | 15.93 <sup>d</sup> (23.52)         |  |  |
| CD (p=0.05) |                          |                                  | 0.05                               | Control     | $0.00^{\mathrm{f}}$      | 00.00 <sup>f</sup> (00.44)         |  |  |
|             |                          |                                  |                                    | CD (p=0.05) |                          | 0.10                               |  |  |

\*Values are mean of three replications

Values in parentheses are arcsine transformed values

In a column, means followed by a common letter are not significantly different at the 5% level by DMRT.

Changes in the initial external pH affect many cellular processes *viz.*, regulation and biosynthesis of secondary metabolites (Reddy *et al.*, 2011). Optimum pH for antibiotic production in *Streptomyces* cultures has been reported to be near neutral (Oskay *et al.*, 2004). In this investigation the optimum pH for the maximum production of antibiotics was found to be pH 7.5. The actinobacteria S. *aureus* strain BG03 cultivated at this pH showed the highest antifungal activity against *R. bataticola* compared to other pH level. Similar findings were earlier reported by Singh *et al.* (2017) who observed maximum antibiotic production in *S. chilikensis* ACITM-1 at pH 7.5.

| Table 5. Antimicrobial compounds identified in the ethyl acetat | e extract of S. aureus strain BG03 by GC- |
|---|---|
| MS analysis   |   |

| R. Time | Compounds  | Molecular<br>formula                           | Compound nature            | Area % | Activity                                | References                          |
|---------|--|--|----------------------------|--------|---|-------------------------------------|
| 18.79   | 9,12- Octadecadienoic acid<br>(Z,Z)- methyl ester                        | $C_{19}H_{34}O_2$                              | Linoleic acid              | 40.47  | Antifungal,<br>Nematicide               | Yu <i>et al.</i> , 2005             |
| 18.98   | 6- Octadecenoic acid, methy ester,(Z)                                    | $C_{19}H_{36}O_2$                              | Stearic acid               | 1.41   | Antifungal                              | Rahuman <i>et al.</i> , 2000        |
| 21.06   | 1h-1, 2, 4- triazole – 1-<br>ethanol, alpha-butyl                        | C14H17Cl2N3O                                   | Alcohol                    | 4.32   | Antifungal,<br>antimicrobial.           | Chandrasekaran <i>et al.</i> , 2011 |
| 14.10   | Pyrrolo {1,2-a} pyrazine-<br>1,4-dione, hexahydro-3<br>(2-methylpropyl)- | $C_{11}H_{18}N_2O_2$                           | Alkaloid                   | 1.34   | Antimicrobial,<br>anti-<br>inflammatory | Yassa et al., 2009                  |
| 11.52   | 1- Nonadecene  | C <sub>19</sub> H <sub>38</sub>                | Alkene<br>compound         | 0.85   | Anti-fungal activity                    | Dalli <i>et al.</i> , 2007          |
| 5.88    | Undecane   | $C_{11}H_{24}$                                 | Alkene<br>compound         | 0.11   | Anti-fungal<br>activity                 | Dalli <i>et al.</i> , 2007          |
| 11.96   | Pentadecanoic acid   | $C_{15}H_{30}O_{2}$                            | Fatty acid                 | 0.33   | Antibacterial,<br>Antifungal            | Aparna <i>et al.,</i><br>2012       |
| 19.18   | 9-Octadecenoic acid (z)-,<br>methyl ester                                | C <sub>19</sub> H <sub>36</sub> O <sub>2</sub> | Oleic acid<br>methyl ester | 1.00   | Antimicrobial<br>Nematicidal            | Yu <i>et al.</i> , 2005             |
| 9.18    | Cyclooctasiloxane,<br>hexadecamethyl-                                    | $C_{16}H_{48}O_8Si_8$                          | Alkane<br>compound         | 0.21   | Antifungal,<br>Nematicide               | Balogun <i>et al.</i> , 2013        |
| 22.57   | Docosane   | $C_{22}H_{46}$                                 | Acyclic<br>alkanes         | 0.46   | Antimicrobial                           | Ghosh <i>et al.</i> , 2011          |

Praveen et al., (2008) reported that extreme pH was unfavorable for production of antibiotics. The optimum pH for growth and bioactive production in S. *purpurascens* (R3) was reported to be pH 7.0 (Bundale et al., 2015). Ahmed et al., (2016) found that Streptomyces strain (SP 1 and SP 28) showed high antifungal activity at the pH ranged from 7-7.5.

The incubation temperature also was found to have an effect on growth as well as bioactive metabolite production. The culture filtrate of *S. aureus* strain BGO3 extracted from the medium incubated at 35° C showed the highest growth inhibition of *R. bataticola* (46.90 % inhibition and 20.66 mm inhibition zone). The medium incubated at 30° C reduced the mycelial growth of *R. bataticola* by 5.68 per cent. The highest temperature of 40° C showed the lowest growth inhibition of 15.93 mm with inhibition zone of 0.23 mm (Table 4).

It was evident from the present study S. *aureus* strain BG03 showed antifungal action against *R. baticola* at 35° C. The media incubated at 30° C also inhibited the mycelial growth of *R. bataticola* to an extend of 35.68 per cent. Therefore the optimum temperature for the maximum production of bioactive metabolites in S. *aureus* strain BG03 found to be 35° C. The maximum antibiotics production by S. *violates* was observed at 30° C (Hassan *et al.*, 2018). *Streptomyces* sp. KGG32 produced maximum biomass and antimicrobial production at 30° C (Oskay *et al.*, 2004). The optimum growth and antimicrobial compound production of S. *chilikensis* ACITM-1 was reported to be 30° C (Singh *et al.*, 2016).

#### Characterization of antibiotics produced by S. aureus strain BG03

The crude metabolites from the S. *aureus* strain BG03 was analyzed through Gas Chromatography-Mass Spectrometry (GCMS) at Agricultural College and Research Institute, Madurai. A total of seven compounds with antifungal activity were identified from S. *aureus* strain BG03, among these four compound were identified as major compounds *viz.*, 6- Octadecenoic acid, methy ester,(Z), 9,12-Octadecadienoic acid (Z,Z)- methyl ester, 1h-1, 2, 4- triazole-1- ethanol, alpha-butyl and undecane were having maximum probability in the crude metabolite extracts (Table 5)

This results were in agreement with findings of Awla *et al.*, (2016) and Taswar *et al.*, (2017). twenty seven different organic compounds were detected from the crude extract of *Streptomyces* EF37141 through GC– MS analysis. These compounds were antimicrobial and antifungal (Taswar *et al.*, 2017).

# CONCLUSION

From this study, it was concluded that Ken Knight's agar medium is the best medium for growth S. *aureus* strain BG03. The Crawford's media amended with starch as the carbon source and peptone as the nitrogen source with the pH of 7.5 and incubated at the temperature of 35° C were found to be optimum for the maximum production of bioactive compound by S. *aureus* strain BG03.

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