

# Impact of Heat Stress on Physiological traits of Greengram

Rakavi<sup>1\*</sup> B., N. Sritharan<sup>1</sup>, A. Senthil<sup>1</sup>, P. Jeyakumar<sup>1</sup>, S. Kokilavani<sup>2</sup> and S.Pannerselvam<sup>2</sup> <sup>1</sup>Department of Crop Physiology,

<sup>2</sup>Agro Climate Research Centre, Tamil Nadu Agricultural University, Coimbatore

Abiotic stress seriously affect the productivity of legumes worldwide. Among legumes, greengram is very popular and extensively grown in India, however, the production is affected by hostile environments, especially heat stress due to climate change. The global air temperature is predicted to rise by 0.2°C per decade, which will lead to (1.8–4.0°C) higher temperature than the current level by 2100. In the above view, an experiment was conducted in CO 8 green gram by elevating the temperature up to 2°C and 4°C from ambient level to study the effect of elevated temperature on morpho-physiological, biochemical and yield attributes of greengram during different growth stages. The results revealed that when CO 8 greengram exposed to elevated temperature of 4°C from the ambient, showed higher variation in its growth, development and yield attributes when compared with 2°C raise. The impact was lesser when plants subjected to an elevated temperature of 2°C from ambient when compared to 4°C raise. This study helps us to know the response of green gram to different elevated temperature at various crop growth stages.Thus, the present investigation opened new vistas in the field of developmental physiology for identifying the heat tolerant genotypes in greengram.

Key words: Leaf area, Total dry matter, Electrolyte leakage, DPPH assay, Yield traits, Greengram

Pulses are very important in our diet because of its rich protein content and ability to fix atmospheric nitrogen in soils. Among pulses, greengram is very popular and its production is affected by hostile environments, especially elevated temperature due to climate change. The global air temperature is predicted to rise by 0.2°C per decade, which will lead to (1.8–4.0°C) higher temperature than the current level by 2100 (IPCC 2007). Elevated temperature both as a seasonal phenomenon and as a part of climate change, is currently the leading threat to world for the production of crop yields (Darkwab et al., 2016). Suitable climate for cultivation of green gram should be warm, humid and within cardinal temperature range of 25°C to 35°C, with moderate rainfall

The flowering time in green gram is adversely affected by conditions comprising high mean temperature and long day photoperiod. Among the various yield limiting factors, environmental stresses particularly water and temperature play a major role. Pulses are very sensitive to drought, water logging and high temperature.Temperature stress has devastating effects on plant growth and metabolism, as these processes have optimum temperature limits in every plant species. Therefore, it was found that the mechanism of heat stress in plants is important to investigate for future generations (Mahla et al., 2012). With this background the current study is undertaken to know the impacts and effects caused by elevated temperature stress in greengram for designing the future perspectives.

\*Corresponding author's email: rakavi.agri@gmail.com

### **Material and Methods**

The pot culture experiment was conducted during 2017 at the open top chambers located in the Department of Crop Physiology, TNAU, Coimbatore. The greengram variety CO 8 was taken for the experiment. The treatments were ambient temperature that exists under open field condition (T<sub>1</sub>), Elevated temperature of 2°C from the ambient temperature (T<sub>2</sub>), Elevated temperature of 4°C from the ambient temperature (T<sub>3</sub>). The treatments were imposed during Vegetative (S<sub>1</sub>), Flowering (S<sub>2</sub>) and Pod development stages (S<sub>3</sub>). The physiological parameters and yield traits were recorded under elevated temperature stress. The experiment was laid out in Factorial Completely Randomized Design (FCRD) with four replications.

Leaf area per plant was measured using a Leaf area meter (LICOR, Model LI 3000) and expressed as cm<sup>2</sup> plant<sup>-1</sup>. Total dry matter production (TDMP) was estimated by randomly pulling out four plants with intact root system at the end of each stage in each treatment and weighed after drying the plants at 80°C for 48 hour and expressed in mg plant<sup>-1</sup>. Electrolyte leakage was measured by the method described by Bajji et al. (2002) and expressed in per cent. The antioxidant activity to scavenge the stable free radical DPPH was assayed by the method of Brand-William et al. (1995). Different concentration of extracts (20-100 µg/ml) was mixed with 1 ml DPPH solution (0.2 mM/ml in methanol) and incubated at 20°C for 40 minutes in dark condition. After 40 minutes of incubation, the discolourisation of the

purple colour was measured at 518nm. The radical scavenging activity was calculated as follows:

Percentage DPPH	Ac-At			
radical scavenging =	x 100			
	Δ <i>l</i> ·			

Where,

Ac = Absorbance of the control;

At = Absorbance of the test sample and expressed as per cent.

Number of nodules was observed through electron microscope (LEICA model). The number of pods produced in each plant was taken randomly from four plants in each replication from each treatment at the harvest stage. The total pod weight was weighed and expressed in gram plant<sup>-1</sup>. The seed yield was expressed in gram plant<sup>-1</sup>. The data arrived and collected on various parameters from the pot culture experiment were analyzed statistically in FCRD (Factorial Completely Randomized Design) as per the procedure suggested by Gomez and Gomez (1984).

## **Results and Discussion**

#### Leaf area and total dry matter production

Leaf area is a photosynthetic surface area of plants. In our study, leaf area (cm<sup>2</sup> plant<sup>-1</sup>) showed that it was decreased in all the stages viz., vegetative stage (S<sub>4</sub>), flowering stage (S<sub>4</sub>) and pod development stage (S<sub>2</sub>) under elevated temperature stress (Table 1). Higher leaf area was observed in T, (97.29 at S<sub>1</sub>, 134.56 at S<sub>2</sub>, 148.91 at S<sub>3</sub>) when compared with T2 (86.67 at S<sub>1</sub>, 121.40 at S<sub>2</sub>, 132.51 at S<sub>3</sub>) and T<sub>3</sub> (74.93 at S<sub>1</sub>, 104.66 at S<sub>2</sub>, 116.15 at S<sub>3</sub>). Extension of cell wall is decreased by stress and high pH which results in structural changes in the plant cell wall, finally reducing the rate of growth of the stressed plants compared to unstressed plants (Burssens et al., 2000; Pakorn et al., 2009). Total dry matter production (TDMP) is an index of biological output from the net photosynthesis.

Table 1. Effect of elevated temperature on leaf area, total dry matter production (TDMP) and number of nodules

	Leaf Area (cm² plant¹)				TDMP (g plant <sup>-1</sup> )				No. of nodules (plant <sup>-1</sup> )		
Treatments	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean	<b>S</b> <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean	S,	S <sub>2</sub>	Mean
T,	97.29	134.56	148.91	126.91	0.90	1.43	6.10	2.80	8.50	19.00	13.75
Т,	86.67	121.40	132.51	113.52	0.77	1.31	5.30	2.46	5.00	12.50	8.75
T_3	74.93	104.66	116.15	98.57	0.62	0.90	3.99	1.83	1.75	4.50	3.12
Mean	86.29	120.20	132.52	113.00	0.76	1.21	5.12	2.36	5.08	12.00	8.54
		Т	S	T×S		т	S	T×S	т	S	T×S
SEd		2.70	2.70	4.69		0.07	0.07	0.13	0.59	0.48	0.84
CD (P≤0.05)		5.55**	5.55"	9.62 <sup>(NS)</sup>		0.16"	0.16"	0.27**	1.25**	1.02**	1.77°

\*, \*\* and NS denote significance level at P<0.05, P<0.01 and non-significance, respectively.

TDMP significantly differed among the treatments viz., T<sub>1</sub> (ambient condition), T<sub>2</sub> (2°C raise from ambient), T<sub>3</sub> (4°C raise from ambient) and also at different stages (S<sub>1</sub>, S<sub>2</sub> and S<sub>3</sub>). Among all the stages, TDMP (g plant<sup>-1</sup>) was significantly higher in T<sub>1</sub> (0.90 at S<sub>1</sub>, 1.43 at S<sub>2</sub>, 6.10 at S<sub>3</sub>) followed by T<sub>2</sub> (0.77 at S<sub>1</sub>, 1.31 at S<sub>2</sub>, 5.30 at S<sub>3</sub>) and the lowest TDMP was found in T<sub>3</sub> (0.62 at S<sub>1</sub>, 0.90 at S<sub>2</sub>, 3.99 at S<sub>3</sub>). In the aspect of number of nodules, it was found to be peak at flowering stage (S<sub>2</sub>). The number of nodules

per plant recorded in T<sub>1</sub> (8.5 at S<sub>1</sub>, 19.0 at S<sub>2</sub>) was the highest among the treatments. In T<sub>2</sub> (5.0 at S<sub>1</sub>, 12.5 at S<sub>2</sub>) and T<sub>3</sub> (1.75 at S<sub>1</sub>, 4.50 at S<sub>3</sub>) there was a significant reduction in the nodule numbers per plant (Table 1.). Present study can be supported with Kurdali (1996) who reported that biological nitrogen fixation (BNF) in legumes is very sensitive to elevated temperature and almost ceases beyond 35°C. Basu *et al.* (2016) also stated that nodulation and nitrogen fixation terminates at high temperature.

Table 2. Effect of elevated	temperature on y	vield traits
-----------------------------	------------------	--------------

Treatments -	Total Pod weight (g plant <sup>-1</sup> )				Seed yield (g plant <sup>-1</sup> )				
	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean	
T <sub>1</sub>		4.24				3.67			
T <sub>2</sub>	3.82	2.83	3.24	3.29	3.50	3.00	2.89	3.11	
T <sub>3</sub>	3.27	2.11	2.31	2.56	3.11	2.02	2.47	2.53	
Mean	3.77	3.05	3.26	3.36	3.41	2.89	3.00	3.10	
		Т	S	T×S		Т	S	T×S	
SED		0.18	0.18	0.32		0.12	0.12	0.20	
CD (P≤0.05)		0.38**	0.38*	0.66(NS)		0.24**	0.24*	0.42(NS)	

\*, \*\* and NS denote significance level at P<0.05, P<0.01 and non-significant respectively

#### Electrolyte leakage and DPPH assay

Electrolyte leakage, a measure of electrolyte

diffusion, resulting from elevated temperature induced cell membrane leakage, has been used to screen and evaluate different wheat genotypes for thermal tolerance (Blum and Ebercon, 1981; Saadalla *et al.*, 1990) and other plants including wheat (Blum *et al.*, 2001), cotton (Ashraf *et al.*, 1994). In our data on electrolyte leakage was found to be maximum in  $T_3$  (53.8 at  $S_1$ , 59.3 at  $S_2$ , 62.2 at  $S_3$ ) followed by  $T_2$  (47.8 at  $S_1$ , 53.9 at  $S_2$ , 57.9 at  $S_3$ ) (Fig.1). Minimum leakage was observed in  $T_1$  (46.5 at  $S_1$ , 52.5 at  $S_2$ , 56.2 at  $S_3$ ) when compared with other treatments. Plants exposed to 4° C elevated temperatures from ambient recorded the maximum electrolyte leakage. The results on DPPH radical scavenging activity

showed higher value under elevated temperature and the data was given in Fig.1. It was observed that in T<sub>3</sub> plants (46.28 at S<sub>1</sub>, 52.70 at S<sub>2</sub>, and 49.08 at S<sub>3</sub>) the DPPH assay exhibited maximum scavenging activity. In T<sub>2</sub> (45.50 at S<sub>1</sub>, 50.18 at S<sub>2</sub>, 46.50 at S<sub>3</sub>) there was a decline in scavenging activity was observed than T<sub>3</sub>.

Among the treatments,  $T_1$  (44.23 at  $S_1$ , 47.68 at  $S_2$ , 44.25 at  $S_3$ ) had minimum scavenging activity than other treatments. The DPPH scavenging activity was high in 4°C elevated temperature proving the



 $T_{2}: Elevated temperature of 2°C from the ambient temperature <math display="block">S_{2}: Flowering stage$   $T_{3}: Elevated temperature of 4°C from the ambient temperature S_{3}: Pod development stage$ 

Fig.1 Effect of elevated temperature on electrolyte leakage and DPPH Assay

presence and increase in total anti-oxidants. Present research revealed that the plants grown under 4°C elevated temperature ( $T_3$ ) showed a higher activity of DPPH radical scavenging activity. Thus the plants with higher accumulation of antioxidant enzymes to detoxify reactive oxygen species and mitigate oxidative stress-induced damage under elevated temperature (Shah *et al.*, 2001).

## Yield traits

Among the treatments,  $T_1$  (4.24) recorded maximum pod weight showing its better performance under ambient temperature condition (Table 2). Number of pods was decreased when temperature was elevated ( $T_2$  and  $T_3$ ) from the ambient condition. Irrespective of the stages, plants imposed with elevated temperature have recorded the lowest number of pod weight with a mean value of 2.83 ( $T_2$ ) and 2.11 (T<sub>3</sub>). The data on seed yield depicted in Table 3 showed similar trend like total pod weight. Plants under ambient temperature (T1) showed highest seed yield per plant (3.67). It was observed that  $T_3$  had greater impact on seed yield (Table 2.). In T<sub>2</sub> the per cent reduction over control was 18 at S2, and 21 at  $\rm S_{_3}$  respectively but in  $\rm T_{_3}$  plants (44% at  $\rm S_{_2}$ , 32.6% at S<sub>3</sub>), the reduction was higher. Our results confirmed by the earlier findings of the Wang et al. (2006), Krishnamurthy et al. (2011) and Kumar et al. (2013).

In green gram, the leaf area was drastically reduced when the temperature was elevated to 4°C from the ambient condition. The reduction in the TDMP was visualized in almost all the stressed plants. The per cent reduction was more in flowering stage (37.1%) of plants kept under 4°C elevated temperature condition. The present study also indicates that electrolyte leakage was high at flowering stage (59.3%) followed by pod development stage (62.2%). In case of number of nodules per plant, the per cent reduction was very high in plants exposed to 4°C (T<sub>3</sub>) elevated temperature at vegetative (79.4%) and flowering stage (76.3%) over control. Elevated temperature mainly affected the pod and seed yield. Seed yield per plant was reduced up to 45% over control during flowering stage when the temperature increased 4°C from the ambient. From the present study, it was concluded that, when greengram crop undergoes to an elevated temperature of 4°C from the ambient, showed significant changes in its physiology and yield attributes.

## References

- Ashraf, M., Saeed, M.M. and Qureshi, M.J. 1994. Tolerance to high temperature in cotton (*Gossypium hirsutum* L.) at initial growth stages. *Environ. Exp. Bot.*, 34:275-283.
- Bajji, M., Kinet, J.M. and Lutts, S. 2002. The use of the electrolyte leakage method for assessing cell membrane stability as a water stress tolerance test in durum wheat. *Plant growth Regul.*, **36**:61-70.
- Basu, P.S., Ummed singh, S., Anil kumar, R.K., Praharaj and Shivran. 2016. Climate change and its mitigation strategies in pulses production. *Ind. J. Agr.*, 61:71-82.
- Blum, A. and Ebercon, A. 1981. Cell membrane stability as a measure of drought and heat tolerance in wheat. *Crop Sci.*, **21**:43-47

Blum, A., Klueva, N. and Nguyen, H.T. 2001. Wheat cellular

thermo tolerance is related to yield under heat stress. *Euphytica*, **117**:117–123.

- Brand-Williams, W., Cuvelier, M.E. and Berset. C. 1995. Use of free radical method to evaluate antioxidant activity. *Lebensm. Wiss. Technol.*, **28**:25-30
- Burssens, S., Himanen, K., van de Cotte, B.T., Beeckman, M., Van Montagu, D.,Inze and Nathalie, V. 2000. *Planta*, **211**:632-640.
- Darkwab, K., Ambachewa, D., Mohammed, H., Asfawa, A. and Matthew. W.B. 2016. Evaluation of common bean (*Phaseolus vulgaris* L.) genotypes for drought stress adaptation in Ethiopia. *The Crop Journal*, 4(5):367-376.
- Gomez, K.A. and Gomez. A.A. 1984. Statistical procedures for agricultural research. (2<sup>nd</sup> Ed.) *John Wiley and Sons*, USA. p. 680.
- IPCC, 2007. Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. *In:* Solomon, S., D. Qin, M. Manning, Z. Chen, M. Marquis, K. B. Averyt, M. Tignor and H. L. Miller (Eds). Cambridge University Press, Cambridge, UK, pp 23-27.
- Krishnamurthy, L., Gaur, P.M., Basu, P.S., Chaturvedi, S.K., Tripathi, V., Vadez, A., Rathore, R., Varshney, K. and Gowda, C.L.L. 2011. Large genetic variation for heat tolerance in the reference collection of chickpea (*Cicer arietinum* L.) germplasm. *Plant Gen. Reso.*, **9**:59-61.

- Kumar, N., Nandwal, A.S., Waldia, R.S., Kumar, S., Devi, S., Singh, S. and Bhasker, P. 2013. High Temperature tolerance in chickpea genotypes as evaluated by membrane integrity, heat susceptibility index and chlorophyll fluorescence techniques. *Ind. J. Agrl. Sci.*, 83(4): 467- 471.
- Kurdali, F. 1996. Nitrogen and phosphorus assimilation, mobilization and partitioning in rainfed chickpea (*Cicer* arietinum L.). Field Crops Research, 47: 81–92.
- Mahla, R., Madan, S., Munjal, R. and Behl, R.K. 2012. Heat induced oxidative stress and changes in protein profile in wheat cultivars. Quality Assurance and Safety of Crops and Foods, 4(3):137-137.
- Pakorn, T., Taychasinpitak, T., Jompuk, C. and Jompuk, P. 2009. Effects of Acute and Chronic Gamma irradiations on in vitro culture of Anubias congensis. *J. Nat. Sci.*, 43: 449 -457.
- Sadalla, M.M., Quick, J.S. and Shanahan, J.F. 1990. Heat tolerance in winter wheat. II. Membrane thermostability and field performance. *Crop Sci.*, **30**: 1248-1251.
- Shah, K., Kumar, R.G., Verma, S. and Dubey, R.S. 2001. Effect of cadmium on lipid peroxidation, superoxide anion generation and activities of antioxidant enzymes in growing rice seedlings. *Plant Science*, **161(6)**:1135-1144.
- Wang, J., Gan, Y.T., Clarke, F. and McDonald, C.L. 2006. Response of chickpea yield to high temperature stress during reproductive development. *Crop Sci.*, 46: 2171–2178.